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Patentanmeldung Nr. Patent application No. Demande de brevet n°

03017551.7



Der Präsident des Europäischen Patentamts; Im Auftrag

For the President of the European Patent Office Le Président de l'Office européen des brevets p.o.

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Anmeldung Nr:

Application no.:

03017551.7

Demande no:

Anmeldetag:

Date of filing:

07.08.03

Date de dépôt:

Anmelder/Applicant(s)/Demandeur(s):

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Bezeichnung der Erfindung/Title of the invention/Titre de l'invention: (Falls die Bezeichnung der Erfindung nicht angegeben ist, siehe Beschreibung. If no title is shown please refer to the description.

Si aucun titre n'est indiqué se referer à la description.)

RA antigenic peptides

In Anspruch genommene Prioriät(en) / Priority(ies) claimed /Priorité(s) revendiquée(s)
Staat/Tag/Aktenzeichen/State/Date/File no./Pays/Date/Numéro de dépôt:

Internationale Patentklassifikation/International Patent Classification/Classification internationale des breyets:

C07K14/00

Am Anmeldetag benannte Vertragstaaten/Contracting states designated at date of filing/Etats contractants désignées lors du dépôt:

AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HU IE IT LU MC NL PT RO SE SI SK TR LI

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EPO - Munich 66 **0.7. Aug. 2003**

Case 21796

RA antigenic peptides

The present invention provides novel naturally-processed RA antigenic peptides which are candidate markers for erosive and non-erosive RA. These antigenic peptides are presented by human MHC class II HLA-DR molecules. Moreover, these antigenic peptides linked to MHC class II molecules, as well as antibodies reactive with said antigenic peptides, nucleic acids encoding said antigenic peptides, and nucleic acid constructs and host cells for expressing said antigenic peptides are provided. The antigenic peptides of the invention as well as the polypeptides they are derived from can be used as markers in diagnosis of RA and in therapy as anti-RA vaccines.

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Rheumatoid Arthritis (RA), originally termed chronic polyarthritis, is a systemic autoimmune disease and one of the most debilitating forms of articular inflammation (Feldmann, M. et al., Cell 85 (1996) 307-310; Dedhia, H.V. & DiBartolomeo, A., Critical care clinics 18 (2002) 841-854). Typically, RA causes joint pain, deformities and severe joint stiffness. The disease can also have its manifestation outside the joints, especially in patients who are positive for an autoantibody, termed "rheumatoid factor" (RF) (Mageed, R.A., in: van Venrooij, W.J. & Maini, R.N. eds., Manual of biological markers of disease, Kluwer Academic Publishers (1996) 1-18). RA occurs quite frequently in the Caucasian population with the susceptibility to RA being influenced by genetic and environmental factors. Both have a crucial effect on the onset and the progression of this autoimmune disease. Approximately 4% of the total population has an increased genetic susceptibility to RA, roughly 20% of which (around 1% of the total population) develops RA as a result of, as yet, uncharacterized non-inheritable factors. Beyond that, RA shows a significant bias in the sex ratio: women have a three fold higher risk for RA than men, indicating that sex hormones may also be involved in the pathogenesis.

In the beginning, RA progresses slowly. Typical early stage symptoms are palm sweating, morning stiffness of fingers and symmetrical joint (www.medicine-worldwide.de). In addition, rheumatoid nodules can appear which is an indication for tissue affection outside the joints. In a simplified model, the immune system produces autoantibodies against healthy tissue (www.netdoktor.de). These autoantibodies attack the articular cartilage in the joint leading to its inflammation and later on to its destruction. This destruction stimulates the immune system to produce more autoantibodies. In addition, cytokines like tumor necrosis-factor alpha (TNF-α) and Interleukin-1 (IL-1) are produced which enhance the inflammatory reaction even further (Houssiau, F.A., Clin Rheumatol 14 Suppl 2 (1995) 10-13). The synovium begins to swell due to infiltration of additional cells of the immune system, such as macrophages and T cells. These cells are actively involved in causing further cell death and in driving joint inflammation (Fox, D.A., Arthritis Rheum 40 (1997) 598-609; Choy, E.H. & Panayi, G.S., N Engl J Med 344 (2001) 907-916). This process resembles a vicious circle of autoantibody production, joint inflammation and joint destruction.

Typically, RA progresses chronically, with 85-90% of all RA patients showing a mild to moderate disease development. Aggressive disease forms leading to complete loss of joint function up to the degree of invalidity is experienced by 10-15% of the patients. In this advanced RA state, patients have a permanent articular inflammation and display rheumatoid nodules. They suffer from strong chronical pain and the inflammation leads to severe finger stiffness and irreversible joint deformations or dislocations.

Diagnosis

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There is growing evidence that therapeutic intervention early in the disease can reduce the extent of joint damage (Egsmose, C. et al., J Rheumatol 22 (1995) 2208-2213; Van der Heide, A. et al., Ann Intern Med 124 (1996) 699-707). Since treatment with disease-modifying antirheumatic drugs (DMARDs) is only justified when the risk:benefit or cost:effectiveness ratios are favorable, it is mandatory to be able to differentiate between RA and other forms of arthritis shortly after onset of the disease (Kirwan, J.R. & Quilty, B., Clin Exp Rheumatol 15 (1997) 15-25). The diagnosis is made by established criteria based on clinical history, physical examination and laboratory tests. The American Society of Rheumatism published a catalog of criteria to help gaining objective evidence for RA (Arnett, F.C. et al., Arthritis Rheum 31 (1987) 315-324). But so far, not a single test is available which is specific for RA. Several biological and biochemical markers, e.g. Creactive protein (CRP), erythrocyte sedimentation rate (ESR), antinuclear antibody (ANA) or RF are utilized for the evaluation of RA. However, these markers are non-specific, as they appear in other inflammatory or autoimmune diseases as well. The RF, for instance, is

an autoantibody that is present in the serum of approximately 50% of RA patients. Since increased levels of the same autoantibody can also be found in the context of other inflammatory diseases, such as Sjögren syndrome, endokarditis or chronical hepatitis, RF is unsuitable to serve as a diagnostic marker for RA. Rather than being of diagnostic value per se, the above mentioned biochemical and biological markers are useful for assessing disease activity and prognosis as well as in the treatment and management of RA patients (Nakamura, R.M., J Clin Lab Anal 14 (2000) 305-313).

Recently, a diagnostic set of criteria was developed that consists of clinical and biochemical aspects which were claimed to discriminate, at an early state, between self-limiting, persistent non-erosive, and persistent erosive RA (Visser, H. et al., Arthritis Rheum 46 (2002) 357-365). Self-limiting arthritis was characterized by natural remission: there was no arthritis on examination in a patient for a certain period of time. Erosive arthritis was defined based on the presence of erosions on radiographs of the hands and/or feet. In particular, the use of antibodies recognizing cyclic citrullinated peptides appears to be promising and suggests an important role for citrullinated antigens in the early diagnosis and prognosis of erosive RA (Schellekens, G.A. et al., J Clin Invest 101 (1998) 273-281; Vincent, C. et al., J Rheumatol 25 (1998) 838-846). The early recognition of erosive RA allows early intervention with DMARDs, which will lead to earlier disease control and improvement of disease outcome (Symmons, D.P.M. et al., J Rheumatol 25 (1998) 1072-1077; Anderson, J.J. et al., Arthritis Rheum 43 (2000) 22-29). Likewise, early recognition of self-limiting and non-erosive arthritis will prevent unnecessary treatment with potentially toxic therapeutics (Fries, J.F. et al., Arthritis Rheum 36 (1993) 297-306.

Therapy

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The goal of any anti-rheumatic therapy is to relieve pain in order to ease the activities of every day life. So far, complete healing of RA is not possible, but by applying modern therapies the progression of the disease can be slowed down or even stopped. Due to individual differences, each patient requires an individualized therapy and early diagnosis, as mentioned before, is desirable. RA therapy is complex and includes lifelong medicinal treatment as well as physio- and radiotherapy. DMARDs used in RA therapy are basic therapeutics (e.g. Methotrexate, Sulfasalazin, Hydroxychloroquin, Leflunomid, Azathioprin), cortisone, non-steroidal anti-inflammatory drugs (NSAID) or monoclonal antibodies against the pro-inflammatory cytokines TNF-a, IL-1ß or their respective receptors (http://rheuma-online.de). These drugs have all in common that they are inhibitors of inflammation by suppressing the immune response. The main disadvantage is their lack of specificity for RA, their adverse effects and their inability to effectively target the causes of RA.

Autoimmunity

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Autoimmunity starts when a specific adaptive immune response is initiated against self antigens (autoantigens) manifested by the development of self-reactive T or B cells. The normal consequence of an adaptive immune response against a foreign antigen is the clearance of the antigen from the body. When an adaptive immune response develops against a self antigen, however, the antigen can in most cases not be completely removed from the body, leading to a sustained immune response. As a consequence, the effector mechanisms of immunity cause chronic inflammatory injury to tissues. The mechanisms of tissue damage are essentially the same in autoimmune disease as those that operate in protective immunity and in hypersensitivity. Even though it is not well understood what triggers autoimmunity, several events which are nowadays believed to contribute to the induction of autoimmune diseases and selection of autoantigenic targets have been summarized most recently (Marrack, P. et al., Nat Med 7 (2001) 899-905).

Autoimmune diseases are controlled by properties of particular genes of each individual and environmental factor. The host's genes affect the susceptibility to autoimmunity at least at three levels. First, some of the genes affect the overall reactivity of the immune system and, thus, can predispose the individual to certain or to several different types of autoimmune diseases. Second, this altered immunoreactivity is funneled to particular autoantigens and tissues by other genes that affect recognition of antigenic peptides by T cells. Third, still other genes act on the ability of target tissues to modulate immune attack for instance by influencing the activity of effector cells of the immune system which are destined to initiate an autoaggressive attack. The latter two sets of genes dictate which antigens will be the targets of autoimmunity and hence which organs will be attacked and what damage will occur.

In addition, signals from the environment influence the development of autoimmunity at the same three levels, by affecting the overall reactivity of the immune system, the antigen-specificity and the state of the potential target tissue. And finally, there is cross-talk between genetic and environmental factors.

Major histocompatibility complex (MHC)

Population studies, genotyping and modern approaches at the molecular level have unanimously shown that certain genes encoded by the major histocompatibility complex (MHC) confer a significantly higher risk for the development of RA (Stastny, P., Tissue Antigens 4 (1974) 571-579; Wordsworth, P. et al., PNAS 86 (1989) 10049-10053; Wordsworth, P. & Bell, J., Springer Semin Immunopathol 14 (1992) 59-78). In particular, the class II MHC alleles *HLA-DRB1*0101*, *0401, *0404 and *0405 in several ethnic groups

increase the susceptibility to RA (Reveille, J., Curr Opin Rheumatol 10 (1998) 187-200). E.g. more than 90% of RF-positive RA patients carry one of these susceptibility alleles. HLA class II molecules are MHC-surface proteins that bind antigenic peptides within the cell and present them on the surface of antigen-presenting cells for interaction with the T cell receptors of CD4⁺ helper T lymphocytes, thereby initiating a cellular immune response (Banchereau, J. & Steinman, R.M., Nature 392 (1998) 245-252). The RA-association of particular HLA class II molecules together with the presence of large numbers of activated CD4⁺ T cells in synovial tissue has supported the model of disease induction in which disease-associated HLA-DR molecules present disease-relevant (e.g. synovial) autoantigens and cause stimulation and expansion of synovial T cells, which then drive the inflammatory process (Striebich, C.C. et al., J Immunol 161 (1998) 4428-4436).

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MHC class II HLA-DR (short: DR) proteins are heterodimers consisting of monomorphic α- and extremely polymorphic β-chains that bind peptide antigens in a peptide binding groove. This groove generally has four major pockets to accept side chains at relative positions 1, 4, 6 and 9 of the peptide (Stern, L.J. et al., Nature 368 (1994) 215-221). The allelic variations between HLA class II molecules account for the differential ability to bind antigenic peptides. This is the rationale why individuals differing in their HLA alleles have divergent antigenic peptide repertoirs, thereby leading to differences in the quality of immune responses (Messaoudi, I. et al., Science 298 (2002) 1797-1800).

Peptides bound by class II MHC molecules are typically longer and more heterogeneous in size (11-25 amino acids) than the peptides bound by class I MHC molecules (8-10 amino acids). This difference arises because the peptide binding groove of class II proteins is open and while peptides are gripped in the midle, their ends can extend out of the groove in a variable fashion (Jones, E.Y., Curr Opin Immunol 9 (1997) 75-79). As a consequence, class II molecules typically bind sets of overlapping peptides that share a common core sequence, termed "T cell epitope", but have different lengths.

More than a decade ago, it was recognized that the DR β chains encoded by RA-linked DRB1 alleles, although exhibiting polymorphic differences, all share a stretch of identical or almost identical amino acids at positions 67-74, known as the "shared epitope" (Gregersen, P.K. et al., Arthritis Rheum 30 (1987) 1205-1213). Since immunity to autoantigens has been regarded central to the pathogenesis of RA, it was hypothesized that the shared epitope could impose disease linkage on the respective DR molecules by at least two different mechanisms: first, by selecting the relevant autoantigenic peptides for presentation, and second, by selecting the appropriate autoreactive T cell specificities during ontogeny. The three-dimensional structure of DR molecules has indeed revealed that the shared epitope is located in the center of the α -helix flanking one side of the

peptide binding groove (Stern, L.J. et al., Nature 368 (1994) 215-221). Thus, strategically this shared epitope region is positioned in such a way that it can interact with both bound peptide and T cell receptor.

However, one of the unresolved mysteries in rheumatology research is the question what are the key arthritogenic antigens and epitopes in man that trigger the onset and the development of RA. Although autoantibodies of different specificity have been identified in serum and synovial fluid of patients it is often unclear whether the antigens which were released at the time of cartilage degradation, were initiating pathogenicity or whether they are merely a consequence of antigen spreading as a result of inflammation (Corrigall, V.M. & Panayi G.S., Crit Rev Immunol 22 (2002) 281-293). Furthermore it is difficult to define pathogenic mechanisms in which the antigen is present throughout the body, including the joint, but the pathology is targeted solely or predominately to the joint.

Autoantigens

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The large number of possible autoantigens in RA is derived from studies using sera or, less frequently, T cells from patients with established chronic RA. One of the most convincing joint-specific antigen that has been proposed in the context of DR molecules, is type II collagen (CII), the predominant protein in articular cartilage. Autoantibodies against CII were found in elevated concentrations in the serum and joints of RA patients although it is not yet clear whether anti-CII antibodies are pathogenic in RA (Banerjee, S. et al., Clin Exp Rheumatol 6 (373-380). Snowden and coworkers have shown that peripheral blood T cells from RA patients proliferated to CII, most pronounced in those patients with anti-CII antibodies. However, the response was seen only in 50% of patients (Snowden, N. et al., Rheumatology 40 (1997) 1210-1218). In a mouse model immunization with CII was shown to induce arthritis in mice expressing the class II MHC alleles DRB1*0401 and *0101 (Rosloniec, E.F. et al., J Exp Med 185 (1997) 1113-1122; Rosloniec, E.F. et al., J immunol 160 (1998) 2573-2578). The immunodominant epitope in both *0401 and *0101 transgenic mice was localized to peptides within residues 261-273 of human CII (Fugger, L. et al., Eur J Immunol 26 (1996) 928-933). The same epitope of CII was capable of stimulating a T cell response in RA patients, particularly in the early stages of disease. Synovial fluid T cells were especially responsive (Kim, H.Y. et al., Arthritis Rheum 42 (1999) 2085-2093).

Although other cartilage proteins have been proposed as RA candidate antigens, DR4-binding epitopes have been defined only for human cartilage glycoprotein 39 (HCgp39). This protein is secreted by synovial cells and articular chondrocytes and its expression is upregulated in plasma and joints during inflammation (Vos, K. et al., Ann Rheum Dis 59 (2000) 544-548). Similar to CII, HCgp39 treatment induces arthritis in

mice. In addition a HCgp39 response of peripheral blood T cells from RA patients was detected (Verheijden, G.F. et al., Arthritis Rheum 40 (1997) 1115-1125). The predominant epitope recognized by T cells in DR4 patients was defined between residues 263-275 and identical to the immunodominant epitope found in DRB1*0401-transgenic mice after immunization with native HCgp39 (Cope, A.P. et al., Arthritis Rheum 42 (1999) 1497-1507). Although not disease specific, responses to this peptide did correlate with disease activity in RA patients (Baeten, D. et al., Arthritis Rheum 43 (2000) 1233-1243). Antibodies to HCgp39, however, have also been detected in the sera of patients with inflammatory diseases, such as inflammatory bowel disease and systemic lupus erythematosus (SLE), albeit at a lower level than in RA.

In an attempt to track antigen-specific T cells in RA, soluble peptide-DR4 tetrameric complexes were used to detect synovial CD4⁺ T cells reactive with CII or HCgp39 in DR4⁺ patients (Kotzin, B.L. et al., PNAS 97 (2000) 291-296). The CII-DR4 complex bound in a specific manner to CII peptide-reactive T cell hybridomas, but did not stain a detectable fraction of synovial CD4⁺ cells. Almost similar results were obtained with the HCgp39-DR4 complex suggesting that the major oligoclonal CD4⁺ T cell expansions present in RA joints are not specific for the dominant CII and HCgp39 determinants described above.

In summary, despite some strong indications for a CII and HCgp39 association with RA, the evidence that they are important antigens in RA is scanty. A direct proof that peptides of CII or HCgp39 are presented in a class II MHC-restricted manner by antigen-presenting cells with subsequent stimulation and activation of synovial CD4⁺ T cells is still lacking. Furthermore a major problem of animal models is their unknown relevance to RA as CII-induced arthritis by immunizing rats or mice differs in many respects from RA.

Naturally processed MHC class II-associated peptides

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An alternative strategy to the identification of RA-specific autoantibodies and T cells relies on the sequence analysis of naturally processed peptide antigens bound to MHC class II molecules. With the help of monoclonal antibodies, class II MHC molecules conferring susceptibility to RA can be purified from cognate cells. RA-associated peptide antigens can be acid-eluted from purified HLA class II molecules. The mixture of small peptides can be separated by HPLC and the peptide sequence be determined by Edman sequencing or mass spectrometry. Due to limitations with peptide purification and sequencing techniques, peptide sequences were, as yet, only obtained from MHC molecules that have been isolated from cultured B cell lines or large amounts of tissue, and the analysis was restricted to a few abundant peptides (Kropshofer et al., J.Exp.Med. 175 (1992) 1799-1803; Chicz, R.M. et al., J Exp Med 178 (1993) 27-47). As a result of the development of high-resolution microcapillary HPLC columns and more sensitive mass spectrometers, MHC-bound

peptides can be analyzed more efficiently (Dongre, A.R. et al., Eur J Immunol 31 (2001) 1485-1494; Engelhard, V.H. et al., Mol Immunol 39 (2002) 127-137).

In the present invention a modified peptide isolation and sequencing technique was used to investigate the peptide antigen repertoire of HLA-DR4 molecules derived from autologous dendritic cells (DCs) which were pulsed with serum or synovial fluid derived from RA patients. The main advantage of this innovative approach is the usage of human DCs that are professionals in RA-relevant antigen processing and presentation, instead of using transgenic animal models or artificial B cell lines.

DCs are enriched in rheumatoid synovial fluid and tissue and are derived from circulating immature precursors (Thomas, R. et al., J Immunol 152 (1994) 2613-2623). They are the most potent antigen-presenting cells which express high levels of MHC molecules together with a variety of accessory molecules (Mellman, I. et al., Trends Cell Biol 8 (1998) 231-237). In a most recent study, it was shown that ex vivo differentiated human DCs and macrophages that are phenotypically similar to antigen-presenting cells from RA synovial joints, were capable of generating and presenting immunodominant epitopes from CII and HCgp39 (Tsark, E.C. et al., J Immunol 169 (2002) 6625-6633). DC have the capacity to prime CD4⁺ helper T cells and to effectively activate cytotoxic CD8⁺ T cells (Ridge, T. et al., Nature 393 (1998) 474-478). Thus, peptides bound to MHC class II molecules and presented by DCs play a superior role in the pathogenesis of diseases involving T cell-driven immune responses.

Therefore, the problem posed by the lack of knowledge of MHC class II restricted antigenic peptides for RA is solved by providing novel naturally-processed MHC class II associated RA antigenic peptides and the polypeptides they are derived from as markers for RA.

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The present invention provides novel naturally-processed antigenic peptides which are candidate RA markers in erosive and non-erosive RA. These antigenic peptides are presented by human MHC class II HLA-DR molecules derived from dendritic cells which were pulsed with serum or synovial fluid derived from patients with established erosive or non-erosive RA. The MHC class II antigenic peptide of the invention are comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39, and originate from interferon-γ-inducible lysosomal

thiol reductase, apolipoprotein B-100, inter- \(\alpha\)-trypsin inhibitor heavy chain H4, complement C4, complement C3, SH3 domain-binding glutamic acid-rich-like protein 3, interleukin-4-induced protein 1, hemopexin, and Hsc70-interacting protein. The present invention also provides these antigenic peptides and the proteins they are derived from as markers for erosive and/or non-erosive RA. Moreover, these antigenic peptides linked to MHC class II molecules, as well as antibodies reactive with said antigenic peptides, nucleic acids encoding said antigenic peptides, and nucleic acid constructs, host cells and methods for expressing said antigenic peptides are provided. Further methods are provided for isolating and identifying RA antigenic peptides.

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Fig. 1: Diagram of Dendritic cell (DC)-mediated analysis of tissue samples: Dendritic cells (DCs), the most specialized antigen-presenting cells (APCs), are brought in contact with an antigen source (e.g. synovial fluid) under optimal conditions for antigen uptake and antigen processing. As a control, DCs are cultured under the same conditions in the absence of synovial fluid antigens. After maturation of DCs, antigen-loaded MHC class II molecules are purified and the respective MHC class II-associated antigenic peptides are isolated and identified.

Fig. 2A: ION-TRAP MS Base Peak Chromatogram of MHC class II-associated antigenic peptides that were isolated from dendritic cells pulsed with the serum of a RA patient. The peptides were eluted directly from a RP-C18-HPLC column into the ion trap mass spectrometer for immediate MS/MS identification. The numbers indicate the retention times (upper value) and the molecular masses (lower value) of the most prominent peptide peaks in the mixture at the respective time.

Fig. 2B: ION-TRAP MS spectrum of antigenic peptides at a retention time of 65.4 min. The marked peak was further fragmented and corresponded to a doubly charged peptide ion from the inter-alpha-trypsin inhibitor ITIH4 (cf. table 3).

Fig. 2C: ION-TRAP MS/MS spectrum of the doubly charged peptide ion at m/z 977.1. The fragmentation masses, together with the mass of the parent ion, were searched against a non-redundant human database by using the SEQUEST algorithm. The retrieved sequence MPKNVVFVIDKSGSMSGR (one-letter-code) corresponded to the dominant epitope ITIH4 (271-288) of the inter-alpha-trypsin inhibitor. The positions of the assigned series of N-terminal B-ions and C-terminal Y-ions are marked.

The antigenic peptides of the invention are peptides, which are associated with and presented by MHC molecules and thereby can have the potential to activate or tolerize T cells. Antigenic peptides presented by MHC class II molecules are therefore MHC class II associated or MHC class II antigenic peptides, whereas antigenic peptides presented by MHC class I molecules are MHC class I associated or MHC class I antigenic peptides.

Peptides which are derived from proteins that are encoded in the genome of the body or an APC are denoted as "self-peptides". The main function of self-peptides presented by DCs in the peripheral lymphoid organs is thought to be the induction of T cell tolerance to self-proteins. Tolerance is the failure to respond to an antigen; when that antigen is borne by self tissues, tolerance is called self tolerance.

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Antigens which are derived from an individual's own body are called "self antigens" or "autoantigens". An adaptive immune response directed against self antigens is called an autoimmune response. Likewise, adaptive immunity specific for self antigens is called autoimmunity. Autoreactivity describes immune responses directed against self antigens. RA is probably due to an autoimmune response that is based on the involvement of autoreactive T cells and/ or autoreactive antibodies. Immunogenic peptide includes, but is not limited to, an antigenic peptide capable of causing or stimulating a cellular or humoral immune response. Such peptides may also be reactive with antibodies.

Peptides derived from proteins encoded in the genome of bacteria, viruses or other foreign invaders and which differ from self-proteins are called "foreign antigenic" or "foreign" peptides. They are able to elicit a T cell response against foreign proteins they are derived from.

RA antigenic peptides are self-peptides that function as self antigens and as a consequence of the disease erroneously trigger autoreactivity against self tissues.

The present invention provides a MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39. Preferably, the MHC class II antigenic peptide has a length of less than 26 amino acids, more preferably a length of 11 to 25 amino acids. Even more preferred is the antigenic peptide of the invention with a length of 11 to 19 amino

acids. Most preferred is the antigenic peptide of the invention consisting of the peptide binding motif comprising the four anchor amino acids.

The present invention also provides a MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 49, or (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 49 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 3.

Furthermore, a MHC class II antigenic peptide is provided comprising (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50, or (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50 with additional N-and C-terminal flanking sequences of the corresponding sequence of SEQ ID NO. 5.

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The MHC class II associated novel antigenic peptides of the invention originate from interferon-γ-inducible lysosomal thiol reductase (SEQ ID NOs. 1 to 3), apolipoprotein B-100 (SEQ ID NOs. 4 and 5), inter-α-trypsin inhibitor heavy chain H4 (SEQ ID NOs. 6 to 12), complement C4 (SEQ ID NOs. 13 to 18), complement C3 (SEQ ID NOs. 19 to 23), SH3 domain-binding glutamic acid-rich-like protein 3 (SEQ ID NOs. 24 to 27), interleukin-4-induced protein 1 (SEQ ID NOs. 28 to 30), hemopexin (SEQ ID NOs. 31 to 35), and Hsc70-interacting protein (SEQ ID NOs. 36 to 39).

The single peptide binding groove of MHC class II molecules is about 25 Å long, but in contrast to MHC class I molecules, both sides are open (Stern LJ et al., Nature 1994; 368, 215-221). Thus, naturally processed antigenic peptides eluted from human MHC class II molecules have a minimal length of about 11 residues and attain a maximal length of about 25 residues (Chicz RM et al., J Exp Med 1993; 178, 27-47).

The stability of the MHC-peptide interaction is determined by more than a dozen hydrogen bonds involving the peptide backbone and the complementarity between specificity pockets of the binding groove and appropriately located amino acid side-chains of the peptide. The amino acids of the peptide fitting into the respective pockets were named "anchor" residues. With regard to most HLA-DR alleles, these anchors are located at relative positions P1, P4, P6 and P9. The combination of amino acids at these 4 anchor positions conferring high-stability binding to the respective HLA-DR allelic product and vary from allele to allele. The peptide binding motif is defined herein as the sequence of nine amino acids comprising the four anchor amino acids. The peptide binding motif of the MHC class II antigenic peptide of the invention is depicted in SEQ ID NO. 49 for the peptides derived from interferon-γ-inducible lysosomal thiol reductase (SEQ ID NOs. 1 to 3), in SEQ ID NO. 50 for the peptides derived from apolipoprotein B-100 (SEQ ID NOs. 4

and 5), in SEQ ID NO. 51 for the peptides derived from inter-α-trypsin inhibitor heavy chain H4 (SEQ ID NOs. 6 to 12), in SEQ ID NO. 52 for the peptides derived from complement C4 (SEQ ID NOs. 13 to 18), in SEQ ID NO. 53 for the peptides derived from complement C3 (SEQ ID NOs. 19 to 23), in SEQ ID NO. 54 for the peptides derived from SH3 domain-binding glutamic acid-rich-like protein 3 (SEQ ID NOs. 24 to 27), in SEQ ID NO. 55 for the peptides derived from interleukin-4-induced protein 1 (SEQ ID NOs. 28 to 30), in SEQ ID NO. 56 for the peptides derived from hemopexin (SEQ ID NOs. 31 to 35), and in SEQ ID NO. 57 for the peptides derived from Hsc70-interacting protein (SEQ ID NOs. 36 to 39). The peptide binding motif may also comprise at least one, at least two, at least three, at least four or at least five modifications of the amino acid sequence while still attaining the binding capacity of the non-modified peptide binding motif. Preferably, the modified peptide binding motif comprises at least three of the four anchor amino acids of the non-modified peptide binding motif. The amino acid modification may be a conservative amino acid substitution as described below.

Additional binding energy is provided by hydrogen bonds involving residues in front of the P1 anchor and behind the P9 anchor. In agreement with that, in most naturally processed peptides the nonameric core-region (P1-P9) is N- and C-terminally flanked by 3-4 residues. Hence, the majority of peptides are 15-17-mers. Longer peptides protrude from the groove, thereby allowing access of exopeptidases which are trimming both ends.

Therefore, the MHC class II antigenic peptide of the invention comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N- and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39, preferably comprises additional N- and C-terminal flanking amino acid residues providing additional binding energy.

Preferably, the MHC class II antigenic peptide of the present invention has a binding capacity to the corresponding MHC class II molecule of between one tenth and ten-fold the IC₅₀ of a corresponding peptide selected from the group consisting of SEQ ID NOs. 1 to 39. The binding capacity of a peptide is measured by determining the concentration necessary to reduce binding of a labelled reporter peptide by 50%. This value is called IC₅₀. A MHC class II antigenic peptide of the invention maintains its binding capacity to the relevant HLA class II molecules as long as it attains IC₅₀ values between one tenth and 10-fold the IC₅₀ of the established reference peptides.

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Since peptide trimming occurs in an individual fashion both before and after binding into the peptide binding groove, the occurrence of several truncation variants sharing a common nonameric core region is a common feature of MHC class II-bound peptides. Importantly, it was shown that C- or N-terminal truncation variants of the same epitope can trigger divergent T cell responses (Arnold et al., (2002) J. Immunol. 169, 739-749).

Several parameters can be envisaged that have an influence on the relative abundance of truncation variants of a particular epitope, e.g. the abundance and integrity of the antigen of relevance, antigen-associated proteins, the abundance of proteases, the type of proteases available and the supply with competitive antigens and/or peptides. Since the antigen supply is a major characteristic that may correlate with the origin of a sample, the ratio of particular truncation variants of an epitope can be of diagnostic value.

A peptide of the invention is a peptide which either has no naturally-occurring counterpart (e.g., such as an mutated peptide antigen), or has been isolated, i.e., separated or purified from components which naturally accompany it, e.g., in tissues such as pancreas, liver, spleen, ovary, testis, muscle, joint tissue, neural tissue, gastrointestinal tissue, or body fluids such as blood, serum, synovial fluid or urine. Typically, the peptide is considered "isolated" when a preparation comprising a peptide of the invention consists to at least 70%, by dry weight of said peptide and to less than 30% of the proteins and naturally-occurring organic molecules with which it is naturally associated. Preferably, a preparation of a peptide of the invention consists of at least 80%, more preferably at least 90%, and most preferably at least 99%, by dry weight, the peptide of the invention. Since a peptide that is chemically synthesized is, by its nature, separated from the components that naturally accompany it, the synthetic peptide is "isolated".

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The invention further provides analogs of the antigenic peptide of the invention. The term analog includes any peptide which displays the functional aspects of these antigenic peptides comprising the binding capacity IC₅₀ and the recognition by antibodies and cells of the immune system. Analogs exhibit essentially the same IC₅₀ as the corresponding reference peptide. The term analog also includes conservative substitutions or chemical derivatives of the peptides.

The term "analog" includes any polypeptide having an amino acid residue sequence substantially identical to the sequences described herein in which one or more residues have been conservatively substituted with a functionally similar residue and which displays the functional aspects of the peptides as described herein. Examples of conservative substitutions include the substitution of one non-polar (hydrophobic) residue such as phenylalanine, tyrosine, isoleucine, valine, leucine or methionine for another, the substitution of one polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, between threonine and serine, the substitution of one basic residue such as lysine, arginine or histidine for another, or the substitution of one acidic residue, such as aspartic acid or glutamic acid for another.

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The phrase "conservative substitution" also includes the use of a chemically derivatized amino acid in place of a non-derivatized amino acid. "Chemical derivative" refers to a subject polypeptide having one or more amino acids chemically derivatized by reaction of a functional side group. Examples of such derivatized molecules include for example, those molecules in which free amino groups have been derivatized to form amine hydrochlorides, p-toluene sulfonyl groups, carbobenzoxy groups, t-butyloxycarbonyl groups, chloroacetyl groups, acetyl groups or formyl groups. Free carboxyl groups may be derivatized to form salts, methyl and ethyl esters or other types of esters or hydrazides. Free hydroxyl groups may be derivatized to form O-acyl or O-alkyl derivatives. The imidazole nitrogen of histidine may be derivatized to form N-im-benzylhistidine. Also included as chemical derivatives are those proteins or peptides, which contain one or more naturally-occurring amino acid derivative of the twenty standard amino acids. For examples: 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine; and ornithine or citrulline may be substituted for lysine.

The MHC class II antigenic peptides of the invention and the proteins they are derived from can be used as markers in diagnosis of RA and in therapy as anti-RA vaccines. The term marker as used herein refers to a biomolecule, preferably a peptide or a polypeptide, which is expressed in a group of patients with a diagnosed disease, e.g. RA, and attains an abundance that is significantly increased or decreased as compared to a control group.

The marker of the present invention may be used as a prognostic marker to predict the susceptibility to a disease, e.g., to predict the susceptibility to RA, as a diagnostic marker for the diagnosis of a disease, e.g. for the diagnosis of RA, as a differential diagnostic marker to differentiate between different forms of a disease, e.g., to differentiate between different forms of RA, as a prognostic marker for the prediction of the outcome of a disease, e.g., for the prognosis of RA, and as a response marker to determine the efficacy of a therapeutic regime, e.g., as a response marker in the treatment of RA.

In a further embodiment, the MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39 is used as a marker for erosive and/or non-erosive RA.

In a further embodiment, the MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 49, or (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 49 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 3 is used as a marker for non-erosive RA.

In a further embodiment, the MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50, or (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50 with additional N-and C-terminal flanking sequences of the corresponding sequence of SEQ ID NO. 5 is used as a marker for erosive RA.

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In a further embodiment, the MHC class II antigenic peptides of the invention as described above are provided linked to a MHC class II molecule.

Multimers (e.g., dimers, trimers, tetramers, pentamers, hexamers or oligomers) of a class II MHC molecule containing a covalently or non-covalently bound peptide according to the present invention, if conjugated with a detectable label (e.g., a fluorescent moiety, a radionuclide, or an enzyme that catalyzes a reaction resulting in a product that absorbs or emits light of a defined wavelength) can be used to quantify T cells from a subject (e.g., a human patient) bearing cell surface receptors that are specific for, and therefore will bind, such complexes. Relatively high numbers of such T cells are likely to be diagnostic of disease or an indication that the T cells are involved in immunity to the disease. In addition, continuous monitoring of the relative numbers of multimer-binding T cells can be useful in establishing the course of a disease or the efficacy of therapy. Such assays have been developed using tetramers of class I MHC molecules containing an HIV-1-derived or an influenza virus-15 derived peptide (Altman et al. (1996), Science 274:94-96; Ogg et al.

(1998), Science 279:2103- 21061), and corresponding class II MHC multimers would be expected to be similarly useful. Such complexes could be produced by chemical cross-linking of purified class II MHC molecules assembled in the presence of a peptide of interest or by modification of already established recombinant techniques for the production of class II MHC molecules containing a single defined peptide (Kazono et al. (1994), Nature 369:151-154; Gauthier et al. (1998), Proc. Natl. Acad. Sci. U.S.A. 95:11828-118331). The class II MHC molecule monomers of such multimers can be native molecules composed of full-length alpha and beta chains. Alternatively, they can be molecules containing either the extracellular domains of the alpha and beta chains or the alpha and beta chain domains that form the "walls" and "floor" of the peptide-binding cleft.

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The invention also relates to an antibody, fragments or derivatives thereof, directed to and reactive with the above-described MHC class II antigenic peptides. The general methodology for producing antibodies is well known and is disclosed per example in Kohler and Milstein, 1975, Nature 256,494 or in J. G. R. Hurrel, Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press Inc., Boco Raron, FL (1982). The antibodies can be polyclonal or, preferably, monoclonal, or antibody fragments like be F (ab') 2, Fab, Fv or scFv. The antibodies of the present invention may also be humanized (Merluzzi S. et al., (2000), Adv. Clin. Path., 4(2): 77-85) or human antibodies (Aujame L. et al., Hum. Antibodies, (1997), 8(4): 155-168).

The present invention also provides a nucleic acid molecule encoding a MHC class II antigenic peptide of the invention comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N- and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39. Preferably, the nucleic acid molecule is a DNA molecule.

Furthermore, a nucleic acid molecule is provided encoding a MHC class II antigenic peptide of the invention linked to a MHC class II molecule.

This invention also provides a recombinant nucleic acid construct comprising the nucleic acid molecules as described above, operably linked to an expression vector. Expression vectors suitable for use in the present invention comprise at least one expression control element operably linked to the nucleic acid sequence encoding the antigenic peptide or the antigenic peptide linked to a MHC class II molecule. The recombinant expression construct may be a DNA construct.

The expression control elements are inserted in the vector to control and regulate the expression of the nucleic acid sequence encoding the antigenic peptide of the invention. Examples of expression control elements include, but are not limited to, lac system, operator and promoter regions of phage lambda, yeast promoters and promoters derived from polyoma, adenovirus, retrovirus or SV40. Additional preferred or required operational elements include, but are not limited to, leader sequence, termination codons, polyadenylation signals and any other sequences necessary or preferred for the appropriate transcription and subsequent translation of the nucleic acid sequence in the host system. It will be understood by one skilled in the art that the correct combination of required or 10 preferred expression control elements will depend on the host system chosen. It will further be understood that the expression vector should contain additional elements necessary for the transfer and subsequent replication of the expression vector containing the nucleic acid sequence in the host system. Examples of such elements include, but are not limited to, origins of replication and selectable markers. It will further be understood by one skilled in the art that such vectors are easily constructed using conventional methods (www.cellbio.com/protocols.html) or are commercially available.

Another aspect of this invention relates to a host organism or a host cell into which a recombinant nucleic acid construct comprising the nucleic acid molecules as described above, operably linked to an expression vector, has been inserted. The host cells transformed with the nucleic acid constructs encompassed by this invention include eukaryotes, such as animal, plant, insect and yeast cells and prokaryotes, such as E. coli. The means by which the nucleic acid construct carrying the nucleic acid sequence may be introduced into the cell include, but are not limited to, microinjection, electroporation, transduction, or transfection using DEAE-dextran, lipofection, calcium phosphate or other procedures known to one skilled in the art (Sambrook et al. (1989) in "Molecular Cloning. A Laboratory Manual", Cold Spring Harbor Press, Plainview, New York).

In a preferred embodiment, eukaryotic expression vectors that function in eukaryotic cells are used. Examples of such vectors include, but are not limited to, retroviral vectors, vaccinia virus vectors, adenovirus vectors, herpes virus vector, fowl pox virus vector, plasmids, or the baculovirus transfer vectors. Preferred eukaryotic cell lines include, but are not limited to, COS cells, CHO cells, HeLa cells, NIH/3T3 cells, 293 cells (ATCC# CRL15731), T2 cells, dendritic cells, monocytes or Epstein-15 Barr Virus transformed B cells.

An antigenic peptide of the invention can be obtained, for example, by extraction from a natural source (e.g., elution from MHC II molecules); by expression of a recombinant nucleic acid encoding the peptide; or by chemical synthesis. A peptide that is

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produced in a cellular system different from the source from which it naturally originates is "isolated," because it will be separated from components which naturally accompany it. The recombinant peptide expressed by a host organism can be obtained as a crude lysate or can be purified by standard protein purification procedures known in the art which may include differential precipitation, size exclusion chromatography, ion-exchange chromatography, isoelectric focusing, gel electrophoresis, affinity, and immunoaffinity chromatography and the like. The extent of isolation or purity can be measured by any appropriate method, e.g. mass spectrometry or HPLC analysis. The peptides may be prepared synthetically by procedures described in Merrifield, (1986) Science 232: 341-347, and Barany andMerrifield, The Peptides, Gross and Meienhofer, eds (N. Y., Academic Press), pp. 1-284 (1979). The synthesis can be carried out in solution or in solid phase or with an automatized synthesizer (Stewart and Young, Solid Phase Peptide Synthesis, 2nd ed., Rockford Ill., Pierce Chemical Co. (1984)).

Therefore, the present invention further provides a method for producing a MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39, comprising the steps of culturing the host cell containing a recombinant nucleic acid construct as described above under conditions allowing expression of said peptide and recovering the peptide from the cells or the culture medium.

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In a further embodiment of the present invention, a method is provided for isolating and identifying MHC class II associated RA antigenic peptides in femtomolar amounts, which method comprises (a) providing immature dendritic cells in a number comprising 0.1 to 5 µg MHC class II molecules; (b) contacting the cells of (a) with serum or synovial fluid and inducing maturation of dendritic cells by adding TNFalpha; (c) isolating class II MHC molecule-antigenic peptide complexes from the cells with methods comprising solubilization of the cells and sequestration of the complexes of MHC class II molecules with antigenic peptides by immunoprecipitation or immunoaffinity chromatography; (d) washing the sequestered complexes of MHC class II molecules with antigenic peptides with water in an ultrafiltration tube; (e) eluting the associated antigenic peptides from the MHC with molecules at 37°C diluted trifluoro (f) separating, detecting and identifying the isolated peptides by liquid chromatography and mass spectrometry. Furthermore, in step (f) of the method, the liquid chromatography comprises a first linear elution step from the reversed-phase material with a volume sufficient to elute the majority of contaminants prior to peptide elution. Moreover, the method may further comprise (g) analyzing the identified peptides by methods comprising a database and a software developed to perform comparative data analysis across multiple datasets.

The amount of tissue or bodily fluid necessary to obtain e.g. 100 ng MHC class II molecules depends on the number of cells that do express MHC class II and on the expression rate of MHC class II molecules: e.g. 100 ng of MHC class II are equivalent to about 2×10^5 mature DCs or 5 to 10×10^6 peripheral blood monocytes or about 5×10^7 peripheral blood mononuclear cells which can be obtained from about 50 ml of blood.

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For the purification of class II MHC molecule-antigenic peptide complexes from cells or tissue, the membranes of the cells or tissue have to be solubilized. Cell lysis may be carried out with methods known in the art, e.g. freeze-and-thaw cycles and the use of detergents, and combinations thereof. Preferred lysis methods are solubilization using detergents, preferably TX-100, NP40, n-octylglucoside, Zwittergent, Lubrol, CHAPS, most preferably TX-100 or Zwittergent 3-12. Cell debris and nuclei have to be removed from cell lysates containing the solubilized receptor-peptide complexes by centrifugation. Therefore, the complexes of class II MHC molecules with antigenic peptides are isolated from the cells with methods comprising solubilization with a detergent.

Furthermore, the MHC class II molecule-peptide complexes are purified from cell lysates by methods comprising immunoprecipitation or immunoaffinity chromatography. For the immunoprecipitation or immunoaffinity chromatography, antibodies specific for MHC class II molecules and suitable for these methods are used. The specific antibodies are preferably monoclonal antibodies, and are covalently or non-covalently e.g. via Protein A, coupled to beads, e.g. sepharose or agarose beads. A selection of the broad panel of antiart comprises: the prior antibodies in HLA anti-HLA-DR antibodies: L243, TU36, DA6.147, preferably L243; anti-HLA-DQ antibodies: SPVL3, TU22, TU169, preferably TU22 and TU169; anti-HLA-DP antibody B7/21 and anti-HLA-A,B,C antibodies W6/32 and B9.12.

Monoclonal antibodies specific for different MHC class II molecules may be commercially obtained (e.g. Pharmingen, Dianova) or purified from the supernatant of the respective hybridoma cells using Protein A- or Protein G- affinity chromatography. Purified monoclonal antibodies may be coupled by various methods known in the art, preferably by covalently coupling antibody amino groups to CNBr-activated sepharose.

Immunoisolation of MHC molecules may be performed by incubating the antibody-beads with the cell lysate under rotation for several hours or chromatographically by pumping the cell lysate through a micro-column. Washing of the antibody-beads may be performed in eppendorf tubes or in the microcolumn. The efficacy of the immunoprecipitation may be analysed by SDS-PAGE and western blotting using antibodies recognizing denatured MHC molecules (anti-HLA-DRalpha: 1B5; anti-HLA class I: HC10 or HCA2).

The sequestered MHC class II molecule-peptide complexes are washed with water or low-salt buffer before elution in order to remove residual detergent contaminants. The low salt buffer may be a Tris, phosphate or acetate buffer in a concentration range of 0.5 – 10 mM, preferably in a concentration of 0.5 mM. In a more preferred embodiment, the MHC class II molecule -peptide complexes are washed with ultrapure water (sequencing grade) conventionally used for HPLC analysis, preferably with ultrapure (sequencing grade) water from MERCK. The washing step may be carried out by ultrafiltration. The ultrafiltration may be carried out in an ultrafiltration tube with a cut-off of 30 kD, 20 kD, 10 kD or 5 kD, preferably of 30 kD and a tube volume of 0.5 – 1.0 ml ("Ultrafree" tubes; Millipore). The washing in the ultrafiltration tube may be carried out 4 to 12 times, preferably 6 to 10 times, with a volume of 10 to 20 times the volume of the beads carrying the receptor-peptide complexes, preferably with a volume of 15 times the beads. The eluted peptides may be separated from the remaining MHC class II molecules using the same ultrafiltration tube. The eluted peptides may then be lyophilized.

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By eluting the peptides from the MHC class II molecules, a complex mixture of naturally processed peptides derived from the source of potential antigen and from polypeptides of intra- or extracellular origin, is obtained. Only after elution, peptides can be separated and subjected to sequence analysis.

The antigenic peptides in the method of the present invention may be eluted by a variety of methods known in the art, preferably by using diluted acid, e.g., diluted acetonitrile (Jardetzky TS et al., Nature 1991 353, 326-329), diluted acetic acid and heating (Rudensky AY et al., Nature 1991, 353, 622-626; Chicz RM et al., Nature 1992, 358, 764-768) or diluted trifluoro acetic acid at 37°C (Kropshofer H et al., J Exp Med 1992, 175, 1799-1803). Most preferably, the peptides are eluted at 37°C with diluted trifluoro acetic acid.

The isolated antigenic peptides are then separated, detected and identified. By detecting it is understood that the amino acid sequence of the individual peptides in the mixture of isolated antigenic peptides is elucidated by methods adequate to detect and sequence femtomolar amounts of peptides. By identifying it is understood that it is

established from which proteins or polypeptides the antigenic peptides are derived and which sequence they constitute within these proteins or polypeptides.

In a first step, the complex mixture of eluted peptides may be separated by one of a variety of possible chromatographic methods, e.g. by reversed phase, anion exchange, cation exchange chromatography or a combination thereof. Preferably, the separation is performed by C18-reverse phase chromatography or by reversed-phase / cation exchange two-dimensional HPLC, denoted as MudPit (Washburn MP et al., Nat Biotechnol., (2001), 19, 242-247).

The separation is done in a HPLC mode utilizing fused-silica micro-capillary columns which are either connected to a nano-flow electrospray source of a mass spectrometer or to a micro-fractionation device which spots the fractions onto a plate for MALDI analysis.

Liquid chromatography comprises peptide fractionation by the use of a strong ion exchange material and a hydrophobic reversed-phase material. For the elution of the peptides from the ion exchange and reversed-phase material different elution programs are run one after another comprising elutions with salt and with organic solvents, e.g., acetonitrile. The elution from the reversed-phase material is conducted in several steps of linear gradients of different lengths and slopes. A contamination in the sample to be fractionated may be any contamination whose elution competes with the detection of the peptide peaks in the mass spectrometer. Therefore, in order to prevent simultaneous elution, the contaminants have to be eluted with a sufficient solvent volume prior to the peptide elution step. Depending on the column used for liquid chromatography the solvent volume sufficient to elute the contaminants prior to the peptide elution step may be 100 to 200 times the column volume.

A variety of mass spectrometric techniques are suitable, preferably MALDI-post source decay (PSD) MS or electrospray ionization tandem mass spectrometry (ESI-MS), most preferably ion-trap ESI-MS.

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The sequences of the individual peptides can be determined by means known in the art. Preferably, sequence analysis is performed by fragmentation of the peptides and computer-assisted interpretation of the fragment spectra using algorithms, e.g. MASCOT or SEQUEST. Both computer algorithms use protein and nucleotide sequence databases to perform cross-correlation analyses of experimental and theoretically generated tandem mass spectra. This allows automated high through-put sequence analysis.

The isolated and identified antigenic peptides of the invention can be validated by the MHC binding motif, the MHC binding capacity and/or by T cell recognition.

MHC binding motif

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Peptides associated to a particular MHC molecule (allelic variant) have common structural characteristics, denoted as binding motifs, necessary to form stable complexes with MHC molecules. Peptide ligands eluted from MHC class I molecules are relatively short, ranging from 8-11 amino acids. Moreover, 2 or 3 side chains of the peptide are relevant for binding. The position of the respective amino acid side chains varies with the HLA allele, most often two of these so-called "anchor" residues are located at positions 2 and 9. With respect to a particular anchor position, only 1 or 2 amino acids normally can function as anchor amino acids e.g. leucine or valine V at position 2 in the case of HLA-A2.

In the case of MHC class II molecules, the peptide length varies from 11 to 25 amino acids, as longer peptides can bind since both ends of the peptide binding groove are open. Most HLA class II molecules accommodate up to 4 anchor residues at relative positions P1, P4, P6 and P9 contained in a nonameric core region. This core region, however, can have variable distance from the N-terminus of the peptide. In the majority of cases, 2-4 N-terminal residues precede the core region. Hence, the P1 anchor residues is located at positions 3, 4 or 5 in most HLA class II associated peptides. Peptides eluted from HLA-DR class II molecules share a big hydrophobic P1 anchor, represented by tyrosine, phenylalanine, tryptophane, methionine, leucine, isoleucine or valine.

The position and the exact type of anchor residues constitute the peptide binding motif which is known for most of the frequently occurring HLA class II allelic products. A computer algorithm allowing motif validation in peptide sequences is "Tepitope", available by vaccinome (www.vaccinome.com).

MHC binding capacity

Peptides identified by the method of the invention may be tested for their ability to bind to the appropriate MHC class II molecule by methods known in the art using, for example, isolated MHC class II molecules and synthetic peptides with amino acid sequences identical to those identified by the method of the invention (Kropshofer H et al., J. Exp. Med. 1992; 175, 1799-1803; Vogt AB et al., J. Immunol. 1994; 153, 1665-1673; Sloan VS et al., Nature 1995; 375, 802-806). Alternatively, a cellular binding assay using MHC class II expressing cell lines and biotinylated peptides can be used to verify the identified epitope (Arndt SO et al., EMBO J., 2000; 19, 1241-1251)

In both assays, the relative binding capacity of a peptide is measured by determining the concentration necessary to reduce binding of a labelled reporter peptide by 50%. This value is called IC₅₀. Peptide binding with a reasonable affinity to the relevant HLA class II molecules attain IC₅₀ values not exceeding 10-fold the IC₅₀ of established reference peptides.

The same binding assays can also be used to test the ability of peptides to bind to alternative class II MHC molecules, i.e., class II MHC molecules other than those from which they were eluted using the method of the invention. The diagnostic methods of the invention using such peptides and therapeutic methods of the invention, using either the peptides or peptides derived from them, can be applied to subjects expressing such alternative class II MHC molecules.

T cell recognition

The epitope verification procedure may involve testing of peptides identified by the method of the invention for their ability to activate CD4+ T cell populations. Peptides with amino acid sequences either identical to those identified in the present invention or corresponding to a core sequence derived from a nested group of peptides identified in the present invention are synthesized. The synthetic peptides are then tested for their ability to activate CD4+ T cells from (a) test subjects expressing the MHC class II molecule of interest and having at least one symptom of the disease; and (b) control subjects expressing the MHC class II molecule of interest and having no symptoms of the disease. Additional control subjects can be those with symptoms of the disease and not expressing the MHC class II molecule of interest.

In some diseases (e.g., those with an autoimmune component) responsiveness in the CD4+ T cells of test subjects but not in CD4+ T cells of the control subjects described in (b) provides confirmatory evidence that the relevant peptide is an epitope that activates CD4+ T cells that can initiate, promote, or exacerbate the relevant disease. In other diseases (e.g., cancer or infectious diseases without an autoimmune component), a similar pattern of responsiveness and non-responsiveness to that described in the previous sentence would indicate that the relevant peptide is an epitope that activates CD4+ T cells that can mediate immunity to the disease or, at least, a decrease in the symptoms of the disease.

CD4+ T cell responses can be measured by a variety of *in vitro* methods known in the art. For example, whole peripheral blood mononuclear cells (PBMC) can be cultured with and without a candidate synthetic peptide and their proliferative responses measured by, e.g., incorporation of [³H]-thymidine into their DNA. That the proliferating T cells are

CD4+ T cells can be tested by either eliminating CD4+ T cells from the PBMC prior to assay or by adding inhibitory antibodies that bind to the CD4+ molecule on the T cells, thereby inhibiting proliferation of the latter. In both cases, the proliferative response will be inhibited only if CD4+ T cells are the proliferating cells. Alternatively, CD4+ T cells can be purified from PBMC and tested for proliferative responses to the peptides in the presence of APC expressing the appropriate MHC class II molecule. Such APC can be B-lymphocytes, monocytes, macrophages, or dendritic cells, or whole PBMC. APC can also be immortalized cell lines derived from B-lymphocytes, monocytes, macrophages, or dendritic cells. The APC can endogenously express the MHC class II molecule of interest or they can express transfected polynucleotides encoding such molecules. In all cases the APC can, prior to the assay, be rendered non-proliferative by treatment with, e.g., ionizing radiation or mitomycin-C.

As an alternative to measuring cell proliferation, cytokine production by the CD4+ T cells can be measured by procedures known to those in art. Cytokines include, without limitation, interleukin-2 (IL-2), interferon-gamma (IFN-gamma), interleukin-4 (IL-4), TNF-alpha, interleukin-6 (IL-6), interleukin-10 (IL-10), interleukin-12 (IL-12) or TGF-beta. Assays to measure them include, without limitation, ELISA, and bio-assays in which cells responsive to the relevant cytokine are tested for responsiveness (e.g., proliferation) in the presence of a test sample.

Alternatively, cytokine production by CD4+ lymphocytes can be directly visualized by intracellular immunofluorescence staining and flow cytometry.

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Moreover, the MHC class II antigenic peptides of the present invention may be used in the diagnosis of RA. Therefore, a further embodiment of the invention is the use of an antigenic peptide according to the present invention as a marker for RA.

Preferably, a MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39 is used as a marker for RA.

In another embodiment, the antigenic peptides of the invention may be used as response markers to track the efficacy of a therapeutic regime. Essentially, a baseline value for an antigenic peptide can be determined, then a given therapeutic agent is administered,

and the levels of the antigenic peptide are monitored subsequently, whereas a change in the level of the antigenic peptide is indicative of the efficacy of a therapeutic treatment.

Furthermore, the antigenic peptides which are only found in certain stages or phases of a disease, preferably of RA, may be utilized as stage-specific markers. Essentially, the levels of the antigenic peptides which have been linked to a certain disease stage are monitored regularly, thereby providing information about the stage of the disease and its progression.

The invention also includes the use of the polypeptides the RA antigenic peptides are derived from as markers for the diagnosis and monitoring of a disease, preferably of RA, and in particular, of erosive versus non-erosive RA. The rationale for the use of the respective proteins is that DCs reside in most tissues where they capture exogenous antigens via specific receptors and via specialized endocytotic mechanisms (e.g. macropinocytosis) followed by presentation of the processed antigens as peptides on MHC class II molecules. Previous studies have shown that the frequency of a peptide epitope found in the context of MHC class II molecules, e.g. the RA antigenic peptides, in the majority of cases mirrors the abundance of the protein from which this particular peptide was derived from. Therefore, not only the RA antigenc peptides but also the corresponding proteins can serve as markers for RA.

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Therefore, in a further embodiment of the present invention, the use of a polypeptide selected from the group consisting of interferon-gamma-inducible lysosomal thiol reductase (SEQ ID NO: 40), apolipoprotein B-100 (SEQ ID NO: 41), inter-alphatrypsin inhibitor heavy chain H4 (SEQ ID NO: 42), complement C4 (SEQ ID NO: 43), complement C3 (SEQ ID NO: 44), SH3 domain-binding glutamic acid-rich-like protein 3 (SEQ ID NO: 45), interleukin-4-induced protein 1 (SEQ ID NO: 46), hemopexin (SEQ ID NO: 47), Hsc70-interacting protein (SEQ ID NO: 48) as a marker for RA is provided. Preferably, the polypeptide is used as a marker for erosive RA. It is also preferred to use the polypetide as a marker for non-erosive RA. Especially preferred is the use of interleukin-4-induced protein 1 (SEQ ID NO: 46) as a marker for RA. The Fig1 polypeptide has not been known as a marker for RA until now, and is considered as an important candidate marker for RA.

The diagnosis of RA can be made by examining expression and/or composition of a polypeptide or peptide marker for RA, by a variety of methods, including enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. A test sample from an individual is assessed for the presence of an alteration in the expression and/or an alteration in composition of a polypeptide or a

peptide of the present invention. An alteration in expression of a polypeptide or peptide can be, for example, an alteration in the quantitative polypeptide expression (i.e., the amount of polypeptide produced); an alteration in the composition of a polypeptide is an alteration in the qualitative polypeptide expression (e.g., expression of a mutant polypeptide or of a different splicing variant).

Both such alterations (quantitative and qualitative) can also be present. "alteration" in the polypeptide expression or composition, as used herein, refers to an alteration in expression or composition in a test sample, as compared with the expression or composition of the peptide or polypeptide in a control sample. A control sample is a sample that corresponds to the test sample (e.g., is from the same type of cells), and is from an individual who is not affected by RA. An alteration in the expression or composition of the peptide or polypeptide in the test sample, as compared with the control sample, is indicative of RA or a susceptibility to RA. Various means of examining expression or composition of a peptide or polypeptide of the present invention can be used, including spectroscopy, colorimetry, electrophoresis, isoelectric focusing, and immunoassays (e.g., David et al., U.S. Pat. No. 4,376,110) such as immunoblotting (see also Current Protocols in Molecular Biology, particularly chapter 10). For example, in one embodiment, an antibody capable of binding to the polypeptide (e.g., as described above), preferably an antibody with a detectable label, can be used. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')2) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin.

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Western blotting analysis, using an antibody as described above that specifically binds to a peptide or polypeptide of the present invention, may be used to measure the level or amount of a peptide or polypeptide in a test sample and comparing it with the level or amount of the peptide or polypeptide in a control sample. Preferably the peptide or polypeptide in a test sample is measured in a homogenous or a heterogenous immuno assay. A level or amount of the polypeptide in the test sample that is higher or lower than the level or amount of the polypeptide in the control sample, such that the difference is statistically significant, is indicative of an alteration in the expression of the polypeptide, and is diagnostic for a RA or a susceptibility to RA.

Therefore, the present invention also relates to a diagnostic composition comprising an antibody reactive with a MHC class II antigenic peptide of the invention.

In a further embodiment the antigenic peptides of the invention or the proteins they are derived from may be used in the prevention and treatment of a disease, preferably of RA.

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One aspect of the invention is a therapeutic purpose, wherein one or more of the identified antigenic peptides are used to vaccinate patients against RA, preferably against erosive and/or non-erosive RA. In the course of the vaccination the antigenic peptide would induce an antigen-specific T cell tolerance in the patient which would ultimately lead to regression of the disease or to an attenuation of disease development.

A promising strategy to induce specific immune tolerance in future clinical trials is the use of DNA tolerizing vaccines. DNA tolerizing vaccines encoding autoantigens alone were shown to reduce T cell proliferative responses (Ruiz, P. et al., J Immunol 162 (1999) 3336-3341), while DNA tolerizing vaccines co-delivering autoantigen plus IL-4 also induced protective T_H2 responses (Garren, H. et al., Immunity 15 (2001) 15-22). Examples of non-polynucleotide-specific tolerizing therapies under development include protein antigens, naturally processed peptides, altered peptide ligands, other biomolecules, such as DNA, or proteins and peptides containing posttranslational modifications, and antigens delivered orally to induce "oral tolerance" (reviewed in: Robinson, W.H. et al., Clin Immunol 103 (2002) 7-12). A potential adverse effect with regard to tolerizing therapies is the development of autoimmunity.

To this end, the relevant RA antigenic peptides may be directly administered to the patient in an amount sufficient for the peptides to bind to the MHC molecules, and provoke peripheral tolerance of T cells.

Alternatively, the antigenic peptides of the invention may be utilized for the generation of vaccines based on DCs. In this case, autologous DCs derived from patients' monocytes may be pulsed with the relevant peptides or recombinant proteins containing the relevant peptide sequences.

Therefore, the present invention provides a pharmaceutical composition comprising a MHC class II antigenic peptide comprising (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or (b) at least the amino acid sequence of the peptide binding motif selected from the group

consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39, an antibody reactive with said antigenic peptide, or a polypeptide selected from the group consisting of SEQ ID NOs 40 to 48, and optionally a pharmaceutically acceptable excipient, diluent or carrier. The antigenic peptide has to be present in an amount sufficient to tolerize specific lymphocytes. Such an amount will depend on the peptide used, the administration, the severity of the disease to be treated and the general conditions of the patient and will usually range from 1 to 50 mg/ml, for example in case of peptides being loaded on dendritic cells.

An acceptable excipient, diluent or carrier may be phosphate buffered saline for *in vitro* studies and physiological salt solutions for *in vivo* applications.

"Vaccination" herein means both active immunization, i. e. the *in vivo* administration of the peptides to elicit an *in vivo* immune tolerance directly in the patient and passive immunization, i. e. the use of the peptides to tolerize *in vitro* CD4+ T lymphocytes or to stimulate autologous or allogeneic dendritic cells, which are subsequently re-inoculated into the patient.

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The present invention also provides the antigenic peptides, antibodies, nucleic acids, host cells, methods, compositions and uses substantially as herein before described especially with reference to the Examples.

Having now generally described this invention, the same will become better understood by reference to the specific examples, which are included herein for purpose of illustration only and are not intended to be limiting unless otherwise specified, in connection with the following figures.

Examples

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The examples below are illustrated in connection with the figures described above and based on the methodology summarized in Fig 1, as described in the following. Commercially available reagents referred to in the examples, were used according to manufacturer's instructions unless otherwise indicated.

Methodology of the invention

Dendritic cells and culturing

The study was performed with human dendritic cells which were differentiated from monocytes, as described below. Monocytes were purified from human peripheral blood. The blood was taken from healthy donors with the following haplotypes: (1) HLA-DRB1*0401, *03011, (2) ·HLA-DRB1*0401, *0304, (3) HLA-DRB1*0401, *1301, (4) HLA-DRB1*0401, *0701.

All cells were cultured in RPMI 1640 medium (short: RPMI) supplemented with 1 mM Pyruvate, 2 mM Glutamine and 10% heat-inactivated fetal calf serum (Gibco BRL, Rockville, MD).

Isolation of peripheral blood mononuclear cells (PBMCs)

Peripheral blood was obtained from the blood bank in Mannheim, Germany as standard buffy coat preparations from healthy donors. Heparin (200 I.U./ml blood, Liquemine, Roche) was used to prevent clotting. Peripheral blood mononuclear cells (PBMCs) were isolated by centrifugation in LSM® (1.077-1.080 g/ml; ICN, Aurora, OH) at 800g (room temperature) for 30 min. PBMCs were collected from the interphase and washed twice in RPMI containing 20 mM Hepes (500g for 15 min, 300g for 5 min). In order to remove erythrocytes, PBMCs were treated with ALT buffer (140 mM ammonium chloride, 20 mM Tris, pH 7.2) for 3 min at 37°C. PBMCs were washed twice with RPMI containing 20 mM Hepes (200g for 5 min).

Generation of dendritic cells from peripheral blood monocytes.

Monocytes were isolated from PBMCs by positive sorting using anti-CD14 magnetic beads (Miltenyi Biotech, Auburn, CA) according to the manufacturer's protocol. Monocytes were cultured in RPMI supplemented with 1% non-essential amino acids (Gibco, BRL, Rockville, MD), 50 ng/ml recombinant human granulocyte macrophage-colony stimulating factor (GM-CSF; S.A. 1.1x10⁷ U/mg) (Leucomax; Novartis, Basel

Switzerland) and 3 ng/ml recombinant human IL-4 (S.A. 2.9x10⁴ U/μg) (R&D Systems, Minneapolis, MN). Monocytes were seeded at 0.3 x 10⁶/ml in 6-well plates (Costar) for 5 days to obtain immature dendritic cells.

The quality of monocyte-derived immature dendritic cells was routinely monitored by flow-cytometric analysis and assessed to be appropriate when they displayed the following phenotype: CD1a (high), CD3 (neg.), CD14 (low), CD19 (neg.), CD56 (neg.), CD80 (low), CD83 (neg.), CD86 (low) and HLA-DR (high). In contrast, mature dendritic cells (cf. below) display the following phenotype: CD1a (low), CD80 (high), CD83 (high), CD86 (high) and HLA-DR (high). Monoclonal antibodies against CD1a, CD3, CD14, CD19, CD56, CD80, CD83, CD86 as well as the respective isotype controls were purchased from Pharmingen (San Diego, CA).

Exposure of dendritic cells to serum or synovial fluid

Serum and synovial fluid were irradiated for 30 min with 137 Cs (70 TBq). To feed dendritic cells with serum- or synovia-derived antigen, 6 x 10^6 immature dendritic cells were pulsed with either 1 ml serum or 0.6 ml synovial fluid. At the same time maturation of dendritic cells was induced by adding 10 ng/ml recombinant human tumor necrosis factor alpha (TNF α ; S.A. 1.1x 10^5 U/µg). As a control, 6 x 10^6 immature dendritic cells were incubated with TNF α alone.

After 24 hrs in culture, mature dendritic cells were harvested by centrifugation at 300g for 10 min. Cells were washed with PBS and transferred to an eppendorf tube. After centrifugation at 400g for 3 min, the supernatant was completely removed and the cells were frozen at -70°C.

Generation of anti-HLA class II beads

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The anti-HLA-DR monoclonal antibody (mAb) L243 (ATCC, Manassas, VA) was produced by culturing the respective mouse hybridoma cell line. mAb L243 was purified using ProteinA sepharose (Pharmacia, Uppsala, Sweden) and immobilized to CNBractivated sepharose beads (Pharmacia) at a final concentration of 2.5 mg/ml, according to the manufacturer's protocol. L243 beads were stored in PBS containing 0.1% Zwittergent 3-12 (Calbiochem, La Jolla, CA).

Nano-scale purification of HLA-DR- peptide complexes

Pellets of frozen dendritic cells were resuspended in 10-fold volume of ice cold lysis buffer (1% Triton-X-100, 20 mM Tris, pH 7.8, 5 mM MgCl₂, containing protease inhibitors chymostatin, pepstatin, PMSF and leupeptin (Roche, Mannheim, Germany)) and lysed in a horizontal shaker at 1000 rpm, 4°C for 1 h. The cell lysate was cleared from cell debris and nuclei by centrifugation at 10000g, 4°C for 10 min. The lysate was coincubated with L243 beads (5-10 µl L243 beads per 100 µl cell lysate) in a horizontal shaker at 1000 rpm, 4°C for 2 hrs. Immunoprecipitated HLA-DR-peptide complexes bound to L243 beads were sedimented by centrifugation at 1000g, 4°C for 1 min and washed four times with 500 µl 0.1% Zwittergent 3-12 (Calbiochem) in PBS.

The efficacy of depletion of HLA-DR-peptide complexes was monitored by analyzing the respective cell lysates before and after immunoprecipitation and aliquots of the beads by western blotting using the anti-HLA-DRα-specific mAb 1B5 (Adams, T.E. et al., Immunology 50 (1983) 613-624).

Elution of HLA-DR-associated peptides

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HLA-DR-peptide complexes bound to L243 beads were resuspended in 100 μl H₂O (HPLC-grade; Merck, Darmstadt, Germany), transferred to an ultrafiltration tube, Ultrafree MC, 30 kD cut-off (Millipore, Bedford, MA) and washed 10 times with 100 μl H₂O (HPLC-grade) by centrifugation for 1-2 min at 10000g at RT. For eluting the bound peptides, 60 μl 0.1% trifluoracetic acid (Fluka, Buchs, Switzerland) in H₂O (HPLC-grade) was added and incubation was performed for 30 min at 37°C. Eluted peptides were collected in a new eppendorf tube by centrigugation of the Ultrafree unit at 10000g for 3 min at RT and immediately lyophilized in a Speed-VacTM vacuum centrifuge.

Fractionation by two-dimensional nanoflow LC

To perform high-throughput sequencing of complex peptide mixtures, the MudPIT (multidimensional protein identification technology) was used (Washburn, M.P. et al., Nat Biotechnol 19 (2001), 242-247) which is based on liquid chromatographic fractionation followed by mass spectrometric sequence determination.

To this end, lyophilized peptides eluted from HLA molecules were resuspended in a buffer containing 5% (v/v) acetonitrile (ACN), 0.5% (v/v) acetic acid, 0.012% (v/v) heptafluoro butyric acid (HFBA) and 1% (v/v) formic acid. The peptide mixture was fractionated on a fused-silica microcapillary column (100 μ m i.d. \times 375 μ m) generated by a Model P-2000 laser puller (Sutter Instrument Co., Novato, CA). The microcolumn was packed with 3 μ m / C18 reversed-phase material (C18-ACE 3 μ m [ProntoSIL 120-3-C18

ACE-EPS, Leonberg, Germany]) followed by 3 cm of 5 μm cation exchange material (Partisphere SCX; Whatman, Clifton, USA).

A fully automated 8-step gradient separation on a LC Packings UltiMate HPLC (LC Packings, San Francisco, USA) was carried out, using the following buffers: 5% ACN / 0.012% HFBA / 0.5% acetic acid (buffer A), 80% ACN / 0.012% HFBA / 0.5% acetic acid (buffer B), 250 mM ammonium acetate / 5% ACN / 0.012% HFBA / 0.5% acetic acid (buffer C), and 1.5 M ammonium acetate / 5% ACN / 0.012% HFBA / 0.5% acetic acid (buffer D). The first 116 min step consisted of a 75 min gradient from 0 to 40% buffer B followed by a 10 min gradient from 40 to 80% buffer B, a 6 min hold at 80% buffer B and a 10 min equilibration step with 100% buffer A. The next 5 steps (146 min each) were characterized by the following profile: 5 min 100% buffer A, 5 min gradient from 0 to x% buffer C, 5 min 100% buffer A, 30 min gradient from 0 to 10% buffer B, 55 min gradient from 10 to 35% buffer B, 20 min gradient from 35 to 50% buffer B, 10 min gradient from 50 to 80% buffer B; a 6 min hold at 80% buffer B, and a 10 min equilibration step with 100% buffer A. The buffer C percentages (x) in steps 2-6 were as follows: 20, 40, 60, 80, and 90%. The 30 min gradient from 0 to 10% buffer B, which is the first linear elution step from the reversed-phase material, was needed in order to sufficiently separate peptide elution from the elution of a major contaminant (m/z=945) which otherwise would have led to the loss of the more hydrophilic peptide peaks. Step 7 consisted of the following profile: 5 min 100% buffer A, 20 min 100% buffer C, 5 min gradient from 0 to 10% buffer B, 35 min gradient from 10 to 35% buffer B, 50 min gradient from 35 to 50% buffer B, 10 min gradient from 50 to 80% buffer B, a 5 min hold at 80% buffer B and a 10 min equilibration step with 100% buffer A. Step 8 was identical to step 7 with the exception of using buffer D instead of buffer C.

Ion trap MS/MS mass spectrometry

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The HPLC column was directly coupled to a Finnigan LCQ Deca XP Plus ion trap mass spectrometer (Thermo Finnigan, San Jose, USA) equipped with a nano-LC electrospray ionization source. Mass spectrometry in the MS/MS mode was performed according to the manufacturer's protocol. Peptides were identified by the SEQUEST algorithm (U.S. patents 6,017,693 and 5,538,897).

MALDI-TOF mass spectrometry

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Peptides spotted onto an AnchorChip plate were co-cristallized with matrix (5 mg/ml; α-cyano-4-hydroxy-cinnamic acid (Merck, Darmstadt, Germany), 50% acetonitrile, 0.1% trifluoroacetic acid). For qualitative analysis of the whole peptide repertoire, samples were analyzed on an UltraflexTM MALDI-TOF mass spectrometer (Bruker, Bremen, Germany), according to the manufacturer's protocol.

Sequence identification by SEQUEST and differential dataset analysis.

MS/MS fragmentation data were analyzed with the software SEQUEST (Thermo Finnigan, San Jose, USA). From an in-house protein database, which was created based on the public databases Swiss-Prot and TrEMBL, SEQUEST extracted for each spectrum all peptide sequences that corresponded to the molecular mass of the parent ion and measured the degree of similarity between the experimental spectrum and the theoretical, in silico generated, spectrum. Only the top-scoring candidate sequence was listed.

The peptide sequences derived from the SEQUEST analysis and their accompanying information on mass accuracy, scoring parameters and peptide origin were stored in an appropriately designed relational database and further processed. Certain constraints were enforced in order to guarantee the storage of only significant sequences with satisfying SEQUEST scores. The two most important constraints were: (i) keep only those sequences that have a cross correlation coefficient (CC) higher than a certain value and (ii) from the remaining sequences keep those ones which have a predefined delta cross correlation coefficient (Δ CC). For both criteria the minimum chosen values are based on empirical knowledge of interpreting SEQUEST results.

A dataset was defined as the sum of data from a particular set of spectra. The design of database and software allowed queries on a single dataset as well as comparisons of multiple datasets. Such a database and software design enables comparative sample analysis, which is not provided by SEQUEST. For instance, possible queries on a single dataset could provide information on the score distribution among the stored spectra, on the existence of further sequence length variants or common subsequences, or on the protein origin of peptide sequences. Since the occurrence of truncation variants of the same epitope is a general characteristic of class II MHC-bound peptides, the existence of length variants in a dataset provides additional strong evidence for the presence of an epitope in a set of spectra.

The most important feature in the analysis of multiple datasets is the possibility to extract a common subset of sequences that satisfies a given criterion. Such a criterion could

be based on sequence similarity, e.g., within all sequences of a collection of datasets, those sequences were selected that had at least one subsequence in common with any other sequence. Such comparisons across different datasets constitute the differential approach (RA samples versus control samples) and thereby optimize the search for candidate RA marker peptides.

The pairwise similarity scores between sequences were calculated by a software routine, which is an implementation of a standard string-comparison algorithm. Subsequently, these scores were used to group closely-related sequences (sequences sharing a common subsequence) in well-separated clusters by an additionally developed software routine, which is based on a well-established algorithm (hierarchical clustering, UPGMA).

The generated clusters (e.g. of peptide truncation variants) were then used to identify closely-related sequences across different datasets.

Overall, the data evaluation software provided the ability to perform swiftly and reproducibly the following:

- -Select from the sequence output generated by SEQUEST those sequences that satisfy reliable empirical criteria.
 - Store the data in a database appropriately designed for the discovery process at hand.
- -Extract information about the sequence content of each stored dataset. This information is valuable in assessing the importance of individual sequences within the given dataset and, consequently, across multiple datasets.
 - -Provide, by virtue of the multiple dataset comparisons, a tool that realizes the differential approach, namely the study of the actual sequence content of one sample versus other(s).

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Example 1

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In this example, the technique mentioned in Figure 1 was used to identify novel HLA-DR-associated peptide markers derived from serum and synovial fluid of patients with non-erosive RA.

 6×10^6 immature dendritic cells were pulsed with either 1 ml serum (3 samples) or 0.6 ml synovial fluid (2 samples) of patients with non-erosive RA and cultured for 24 hrs in the presence of 10 ng/ml TNF α . As a control, 6×10^6 dendritic cells were cultured in the presence of TNF α (10 ng/ml) without adding serum but 1 ml of PBS. In an additional experiment 6×10^6 dendritic cells were pulsed with 1 ml serum from 2 healthy test persons and cultured for 24 hrs in the presence of TNF α (10 ng/ml).

Dendritic cells were lysed in detergent TX-100 and HLA-DR molecules were isolated using mAb L243. HLA-DR-associated peptides were eluted with 0.1% TFA and analyzed by high-throughput 2D-LC-MS/MS technology. Peptide identification was achieved by using the SEQUEST algorithm. The peptide sequences derived from the SEQUEST analysis and accompanying information on mass accuracy, scoring parameters and peptide origin were stored in a database and further processed.

The peptide sequences identified from unpulsed DCs (control 1) and from DCs pulsed with the serum of healthy test persons (control 2) were compared with the peptide sequences identified from DCs pulsed with the serum of non-erosive RA patients. Among the RA-specific sequences, only those peptides were selected for further evaluation that reoccurred in at least three of five non-erosive RA samples.

In each serum sample roughly 600 ± 150 individual peptide sequences (cross correlation coefficient CC > 3.0 and Δ CC > 0.15) were identified. In the synovia samples the number of individual peptide sequences was slightly smaller (400 ± 30). Approximately 80-85% of the peptides found in RA samples were also identified in control samples, underlining the high reproducibility of the analysis. In the majority of cases, several length variants of the same epitope could be identified which is a typical characteristic of class II MHC-bound antigens and supports the validity of the results (Jones, E.Y., Curr Opin Immunol 9 (1997) 75-79). Further confidence in the quality of the data relies on the fact that several of the identified peptides or proteins have already been described in the context of MHC class II molecules: epitopes derived from ubiquitous proteins like Hsp70, enolase, annexin II, cathepsin C or collagen II, as well as from MHC molecules (HLA-A, -B, -C, -E, -G, and β_2 -microglobulin) and CLIP (Chicz, R.M. et al., J Exp Med 178 (1993) 27-47; Sinigaglia, F. & Hammer, J., Curr Opin Immunol 6 (1994) 52-56; Arnold-Schild, D.

et al., J Immunol 162 (1999) 3757-3760; Vogt, A.B. & Kropshofer, H., Trends Biochem Sci 4 (1999) 150-154) were frequently detected.

RA-specific peptide sequences were further validated with regard to binding to the RA susceptibility allele *DRB1*0401* by using the TEPITOPE software (Hammer, J. et al., Adv Immunol 66 (1997) 67-100). This software provides means for the qualitative and quantitative prediction of T cell epitopes.

The output of the study consists of an epitope that occurred, apart from one exception, only in non-erosive RA samples (Table 1).

Interferon-gamma-inducible lysosomal thiol reductase

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A very interesting epitope which was identified in 3 out of 5 non-erosive RA samples from serum and synovia is derived from the interferon-gamma-inducible lysosomal thiol reductase (GILT): the 16-mer GILT (192-207) with the amino acid sequence of SEQ ID NO: 3 (Table 1). Further length variants in three other samples support the relevance of this epitope (Table 1): the 14-mer GILT (192-205; SEQ ID NO: 1) and the 17-mer GILT (192-208; SEQ ID NO: 2).

As judged from the shortest length variant, GILT (192-205), the epitope contains a suitable binding motif, with regard to binding to the RA susceptibility allele *DRB1*0401*: 196M serves as a P1 anchor, 199M as a P4 anchor and 201A as a P6 anchor. According to TEPITOPE scoring, the epitope has a binding score (threshold value) of 1% which is similar to the binding score of an epitope from influenza haemagglutinin (307-319) that was shown to be a strong *DRB1*0401* binder (Table 1) (Rothbard, J.B. et al., Cell 52 (1988) 515-523).

GILT is constitutively expressed in antigen-presenting cells, such as dendritic cells, macrophages and B cells, and facilitates unfolding of endocytosed antigens in MHC class II-containing compartments (MIIC) by enzymatically reducing disulfide bonds (Phan, U.T. et al., J Biol Chem 275 (2000) 25907-25914). Direct binding of GILT to HLA-DR molecules has been reported for B cells (Arunachalam, B. et al., J Immunol 160 (1998) 5797-5806). A rather long second epitope of GILT was found to bind to HLA-DR3 molecules: the 22-mer GILT (38-59)having the amino acid SPLQALDFFGNGPPVNYKTGNL (Chicz, R.M. et al., J Exp Med 178 (1993) 27-47).

In addition to GILT (192-207), another epitope of the same protein was identified in several RA samples, but also in control samples: GILT (210-227) with the amino acid

sequence QPPHEYVPWVTVNGKPLE. This epitope was accompanied by 3 other length variants: the 16-mer GILT (210-225), the 17-mer GILT (210-226) and the 19-mer GILT (210-228).

As indicated by its name, GILT expression can be induced by the pro-inflammatory cytokine interferon gamma (IFN- γ) in various types of cells, including macrophages, endothelial cells and fibroblasts (Luster, A.D. et al., J Biol Chem 263 (1988) 12036-12043). As IFN- γ is known to be present in inflamed joints of RA patients, GILT could become over-expressed in synovia and serum and, hence, could be taken up by DCs as an exogenous antigen. GILT (192-207) may be derived from exogenous GILT. The other GILT epitope, which is also present in the control samples, may be derived from endogenous GILT, expressed by DCs. Alternatively, both GILT (192-207) and GILT (210-227) may be derived from endogenous GILT, in case that GILT processing and GILT-derived epitope presentation by DCs were critically altered upon contact with RA-associated material.

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Example 2

In this example, the same technology was used that has been described in detail in example 1. Serum (4 samples) and synovial fluid (2 samples) of patients with diagnosed erosive RA were utilized in this case to identify candidate markers specific for erosive RA.

The peptide sequences found in the erosive RA samples were compared with the sequences identified in unpulsed DCs (control 1) and in DCs pulsed with the serum of healthy test persons (control 2). Among the RA-specific sequences, only those peptides were selected for further evaluation that re-occurred in at least three of six erosive RA samples.

In this study one epitope was discovered which occurred, apart from one exception, only in erosive RA samples.

Apolipoprotein B-100

The epitope which was mainly found in erosive RA sera (4 out of 6 erosive RA samples) is derived from apolipoprotein B-100: the 16-mer ApoB (2877-2892) with the amino acid sequence of SEQ ID NO: 4 (Table 2). In addition a length variant of the same epitope was identified (Table 2): the 17-mer ApoB (2877-2893; SEQ ID NO: 5). The

following DRB1*0401 binding motif can be predicted: 2881L as a P1 anchor, 2884D as a P4 anchor and 2886N as a P6 anchor (binding score 3%).

In an earlier study on EBV-B cells, the epitope ApoB (2885-2900), which partly overlaps with the epitope described here, has been found in the context of HLA-DR4 (Chicz, R.M. et al., J Exp Med 178 (1993) 27-47).

Apolipoprotein B-100 is a constituent of very low-density lipoproteins (VLDL) and low-density lipoproteins (LDL) and functions as a recognition signal for the cellular binding and internalization of LDL particles by the ApoB/E receptor (Yang, C.Y. et al., Nature 323 (1986) 738-742). Interestingly, an increased ratio of LDL cholesterol to HDL cholesterol was observed among newly diagnosed RA patients (Park, Y.B. et al., J Rheumatol 26 (1999) 1701-1704). The adverse lipid profile in active RA could be improved by treating RA patients with DMARDs without the use of lipid-lowering agents (Park, Y.B. et al., Am J Med 113 (2002) 188-193). Since an increased cardiovascular mortality among patients with chronic inflammatory diseases, such as RA, is well documented (Symmons, D.P. et al., J Rheumatol 25 (1998) 1072-1077) it was suggested that local inflammation in RA leads to altered blood lipid levels, thereby increasing the risk of atherosclerosis. The question whether components of the lipoprotein metabolism are causal for pathogenesis or merely affected by ongoing immune reactions during RA development cannot be answered yet. However, the observation of adverse lipid profiles in RA patients supports the validity of the presented ApoB epitope as a serum-derived RA candidate marker.

The length variant ApoB (2877-2892), but not ApoB (2877-2893), has been identified in samples of two healthy controls (Table 2). Since Apolipoprotein B constitutes 1% of all plasma proteins, the presence of ApoB epitopes in healthy control samples is not surprising. The results suggest that only the length variant ApoB (2877-2893; SEQ ID NO: 5) is specific for erosive RA.

Example 3

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All peptide sequences identified in examples 1 and 2 from non-erosive and erosive RA samples were used in this example to search for common markers relevant for both RA types. The RA-specific sequences were again compared with peptide sequences of the control samples (unpulsed DCs and DCs pulsed with the serum of two healthy test-persons) and only those peptides were selected for further evaluation that re-occurred in at least three of altogether eleven RA samples (erosive and non-erosive RA).

Inter-alpha-trypsin inhibitor

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Six out of seven serum samples (erosive & non-erosive RA) but not any of the controls gave rise to an epitope derived from the heavy chain H4 of the inter-alpha-trypsin inhibitor: ITIH4 (271-287) with the amino acid sequence of SEQ ID NO: 8 (Table 3).

Apart from this major length variant of the ITIH4 epitope, six length variants of the same ITIH4 epitope could be identified (Table 3): the 19-mer ITIH4 (271-289; SEQ ID NO: 6), the 18-mer ITIH4 (271-288; SEQ ID NO: 7), the 16-mer ITIH4 (274-289; SEQ ID NO: 12), the 15-mer ITIH4 (273-287; SEQ ID NO: 10), the 15-mer (274-288; SEQ ID NO: 11) and the 14-mer ITIH4 (274-287; SEQ ID NO: 9).

As judged from the shortest length variant, ITIH4 (274-287), the epitope contains a very strong binding motif, with regard to binding to the RA susceptibility allele DRB1*0401: 277F serves as a P1 anchor, 280D as a P4 anchor and 282S as a P6 anchor (binding score: 1%).

ITIH4 belongs to the Inter-alpha-inhibitor (IaI) family which is a group of serum protease inhibitors that bind to hyaluronic acid (HA) and appear to be involved in acute-phase reactions (Salier, J.P. et al., Biochemical Journal 315 (1996) 1-9).

HA is a polysaccharide found in all tissues of the body, in particular, in loose connective tissue, e.g. joint fluid (Evered, D. & Whelan, J. eds., The Biology of Hyaluronan, John Wiley & Sons (1989)). HA has an important structural function in cartilage and other tissues where it stabilizes the extracellular matrix by forming aggregates with proteoglycans. It has also been assigned important biological functions by regulating cellular activities via binding to cell surface proteins, such as CD44 and ICAM-1 (Knudson, C.B. & Knudson, W., FASEB J 7 (1993) 1233-1241; Hall, C.L. et al., J Cell Biol 126 (1994) 575-588). RA is accompanied by a large increase in total HA in the joint fluid as well as in the serum, suggesting that circulating HA originates from rheumatoid joints (Engström-Laurent, A. et al., Scand J Clin Lab Invest 45 (1985) 497-504).

Complexes of HA and some IoI family members were observed in large amounts in the synovial fluid of RA patients (Jessen, T.E. et al., Biological Chemistry Hoppe-Seyler 375 (1994) 521-526). The role of the IoI-HA complex in inflammatory reactions might be to modify the CD44-HA interaction that mediates leukocyte activation and invasion (Isacke, C.M. & Yarwood, H., Int J Biochem Cell Biol 34 (2002) 718-721). Additionally, synovial fluid of RA patients contains elevated levels of TSG-6, an anti-inflammatory glycoprotein and a member of the hyaladherin family of HA-binding proteins (Wisniewski, H.G. et al., J Immunol 151 (1993) 6593-6601). It has been shown that a complex of TSG-6 with IoI family members inhibits the activity of plasmin, a central molecule in the activation of

inflammation-associated enzymes (Wisniewski, H.G. et al., J Immunol 156 (1996) 1609-1615). A regulation of plasmin activity by several acute-phase plasma proteins, namely TSG-6 and IoI family members, may prove to be important in RA, given the high contents of HA, TSG-6 and IoI family members in synovial fluid of inflamed joints.

This evidence, together with the identification of multiple length variants of the same epitope and a strong HLA-DR4 binding motif, convincingly support the validity of the presented ITIH4 epitope as a serum-derived RA candidate marker.

Complement C4

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In all erosive and non-erosive RA sera tested, another dominant epitope was identified which is derived from complement C4: the 15-mer C4 (1697-1711) with the amino acid sequence of SEQ ID NO: 13 (Table 3). Five additional length variants of the same epitope could be found (Table 3): the 12-mer C4 (1697-1708; SEQ ID NO: 18), the 13-mer C4 (1698-1710; SEQ ID NO: 17), the 14-mer C4 (1697-1710; SEQ ID NO: 15), the 16-mer C4 (1697-1712; SEQ ID NO: 14) and the 18-mer C4 (1697-1714; SEQ ID NO: 16). Moreover, the presented epitope displays a very strong *DRB1*0401* binding motif: 1700Y as P1 anchor, 1704D as P2 anchor and 1706N as P6 anchor (binding score: 1%).

C4 which constitutes approximately 0.5% of plasma protein mass plays a critical role in the triggering of the central pathway of the complement system. The protein is synthesized as a single-chain precursor and, prior to secretion, is enzymatically cleaved to form a trimer of non-identical α -, β -, and γ -chains. The identified epitope C4 (1697-1711) is located at the very C-terminus of the C4 γ -chain. The C4 α -chain is further proteolytically degraded by activated C1 to form the C4a anaphylatoxin, which is a mediator of local inflammatory processes (Moon, K.E. et al., J Biol Chem 256 (1981) 8685-8692).

In general, the complement cascade is involved in the induction and progression of inflammatory reactions and is a major defense system against various pathogenic agents, including bacteria, viruses and other antigens (Morgan, B.P., Methods Mol Biol 150 (2000) 1-13). Inappropriate activation, however, can lead to tissue damage and manifestation of disease (Speth, C. et al., Wien Klin Wochenschr 111 (1999) 378-391).

Activation of the complement system has been repeatedly implicated in the pathogenesis of RA, based on studies showing increased levels of complement metabolites, including C4 and C4a, in plasma, synovial fluid and synovial tissue of RA patients

(Neumann, E. et al., Arthritis Rheum 46 (2002) 934-945). In addition collagen-induced arthritis (CIA) in mice is characterized by the presence of complement activation products (Linton, S.M. & Morgan, B.P., Mol Immunol 36 (1999) 905-914). CIA is prevented after treatment with anti-C5 monoclonal antibodies (Wang, Y. et al., PNAS 92 (1995) 8955-8959) or with soluble CR1, an inhibitor of the complement system, delivered by gene therapy (Dreja, H. et al., Arthritis Rheum 43 (2000) 1698-1709). Activation of complement factors in joints is possibly induced by the presence of various immune complexes and it was hypothesized that stimulation of the innate immune system by infectious agents and cytokines may contribute to the initiation of RA (Friese, M.A. et al., Clin Exp Immunol 121 (2000) 406-414).

Two of the six presented C4 epitopes, the 15- and the 18-mer, were also identified in healthy control samples (Table 3) indicating that only some length variants of this C4 epitope are RA-specific, namely the antigenic peptides of SEQ ID NOs: 14, 15, 17, and 18.

Complement C3

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Another epitope that was found in erosive and non-erosive RA samples is derived from complement C3: the 14-mer C3 (1431-1444) with the amino acid sequence of SEQ ID NO: 21 (Table 3). Four additional length variants of the same epitope were identified in serum (Table 3): the 13-mer C3 (1431-1443; SEQ ID NO: 23), the 15-mer C3 (1431-1445; SEQ ID NO: 20) and the 19-mer C3 (1426-1444; SEQ ID NO: 19). As judged from the shortest length variant, C3 (1431-1443), a DRB1*0401 binding motif can be postulated: 1434Y serves as a P1 anchor, 1437D as a P4 anchor and 1439A as a P6 anchor.

Complement C3 which constitutes about 1-2% of plasma protein mass plays a central role in the activation of the complement system and belongs to the family of the acute-phase proteins. Its processing by C3 convertase to C3a anaphylatoxin and C3b is the central step in both the classical and alternative complement pathways (Barrington, R. et al., Immunol Rev 180 (2001) 5-15). After activation, C3b can bind covalently, via a reactive thiolester, to cell surface carbohydrates or immune aggregates (Isaac, L. & Isenman, D.E., J Biol Chem 267 (1992) 10062-10069). The identified epitope C3 (1431-1444) is located at the C-terminus of C3b.

As already discussed in the context of complement epitope C4 (1697-1711), there is increasing evidence for an important role of components of the complement cascade in the pathophysiology of RA. The result of this study, in which two major epitopes derived from

complement C3 and C4 were identified in serum of RA patients, underlines the close link between the activated complement system and pathogenesis of RA. This coincidence makes a strong argument for the validity of the presented C3 / C4 epitopes as serum-derived candidate RA markers.

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SH3 domain-binding glutamic acid-rich-like protein 3

Another epitope which was elucidated quite frequently in serum of RA patients (5 out of 7 erosive and non-erosive RA samples), is derived from the SH3 domain-binding glutamic acid-rich-like protein 3 (SH3BGRL3): SH3BGRL3 (15-26) with the amino acid sequence of SEQ ID NO: 25 (Table 3). Three length variants of the same epitope were identified (Table 3): the 14-mer SH3BGRL3 (13-26; SEQ ID NO: 26), the 14-mer SH3BGRL3 (15-28; SEQ ID NO: 27) and the 16-mer SH3BGRL3 (13-28; SEQ ID NO: 24). The *DRB1*0401* binding motif is: 17I as P1 anchor, 20Q as P4 anchor and 22S as P6 anchor (binding score 4%).

SH3BGRL3 is a small 10 kD protein that belongs to the SH3BGR family. The precise function of the protein is unknown but a role as a modulator of glutaredoxin biological activity is postulated (Mazzocco, M. et al., Biochem Biophys Res Commun 285 (2001) 540-545). So far, SH3BGRL3 has not been described in the context of RA.

Interestingly, the analysis elucidated a second epitope of the same protein, which was highly abundant in all RA and control samples: the 16-mer SH3BGRL3 (29-44) with the amino acid sequence DGKRIQYQLVDISQDN. In addition multiple length variants of the same epitope were found in most samples as well. As judged from the shortest length variant, SH3BGRL3 (31-42), the epitope contains almost similar DRB1*0401 anchor residues compared with SH3BGRL3 (15-26): 33I serves as a P1 anchor, 36Q as a P4 anchor and 38V as a P6 anchor (binding score -2). This similarity is reflected by comparable binding scores.

The presence of this second SH3BGRL3 epitope supports the validity of the SH3BGRL3 (15-26) epitope because both peptides are derived from the same protein, however, only one of them, epitope SH3BGRL3 (15-26), appears to be generated in a RAspecific manner. A similar observation has been described already for GILT in example 1.

Among the four SH3BGRL3 length variants the longest variant, SH3BGRL3 (13-28), was also identified in a healthy control sample (Table 3). However, this particular length variant was found only one time, which indicates a significant enrichment of the SH3BGRL3 epitope in the context of RA.

Interleukin-4 (IL-4) induced protein 1

In all the investigated sera and synovial fluids (erosive & non-erosive RA), one highly dominant epitope was identified which is derived from the human homolog of the IL-4 induced protein 1 (Fig1): Fig1 (293-309) with the amino acid sequence of SEQ ID NO: 28 (Table 3). The validity of the epitope was further supported by the presence of additional length variants in several samples (Table 3): the 16-mer Fig1 (293-308; SEQ ID NO: 30) and the 19-mer Fig1 (293-311; SEQ ID NO: 29). Moreover, the amino acid sequence displays a typical *DRB1*0401* binding motif: 299V serves as P1 anchor, 302E as P4 anchor and 304S as P6 anchor (binding score 1%).

Two length variants of the same epitope, Fig1 (293-308) and Fig1 (293-309), were identified in one unpulsed sample and in one healthy control sample as well (Table 3). However, the presence of the Fig1 epitope in all RA samples but not in all of the control samples tested strongly indicates an enrichment in the context of RA.

The human fig1 gene was first identified in IL-4-stimulated B cell cultures (Chu, C.C. & Paul, W.E., PNAS 94 (1997) 2507-2512). The human fig1 resides on chromosome 19q13.3-19q13.4, a region previously identified to be involved in susceptibility to autoimmune diseases, including SLE, arthritis, multiple sclerosis, and insulin-dependent diabetes mellitus (Becker K.G. et al., PNAS 95 (1998) 9979-9984). Since its expression is largely limited to immune tissues and its regulation is dependent on IL-4, a key modulator of the immune response, fig1 is thus an attractive candidate gene for autoimmune disease susceptibility (Chavan, S.S. et al., Biochim Biophys Acta 1576 (2002) 70-80). The HLA-DR4-restricted presentation of a Fig1 epitope provides the first indication that Fig1 protein is produced and possibly involved in the disease development of RA. The Fig1 polypeptide has not been known as a marker for RA until now, and is considered as an important candidate marker for RA.

Hemopexin

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Another RA candidate marker which was frequently identified in serum samples (6 out of 7 samples) and in synovia samples (2 out of 4 samples) (erosive & non-erosive RA) is derived from hemopexin (HPX): HPX (351-367) with the amino acid sequence of SEQ ID NO: 32 (Table 3). Several length variants were found which support the validity of this epitope (Table 3): the 13-mer HPX (351-363; SEQ ID NO: 33), the 14-mer HPX (350-363; SEQ ID NO: 34), the 15-mer HPX (351-365; SEQ ID NO: 35) and the 18-mer HPX (351-368; SEQ ID NO: 31). Furthermore, the epitope contains a very strong DRB1*0401 binding

motif: 355I serves as a P1 anchor, 358D as a P4 anchor and 360V as a P6 anchor (binding score: 1%).

Two length variants of the same epitope, HPX (351-367; SEQ ID NO: 32) and HPX (351-365; SEQ ID NO: 35), could also be identified in healthy control samples (Table 3) indicating that only some length variants are specific for RA, namely the antigenic peptides of SEQ ID NOs. 31, 33 and 34.

HPX is a 60 kD plasma glycoprotein with a high binding affinity to heme (Müller-Eberhard, U., Methods Enzymol 163 (1988) 536-565). It is mainly expressed in the liver, and belongs to the acute-phase proteins the synthesis of which is induced in an inflammatory situation. RA is a chronic inflammatory autoimmune disease and elevated levels of several acute-phase proteins, including C-reactive protein and serum amyloid A, have been reported (Nakamura, R., J Clin Lab Anal 14 (2000) 305-313). HPX is responsive to the cytokines IL-1 and IL-6, which are upregulated in patients suffering from RA (Feldmann, M. & Maini, R.N., Rheumatology 38, Suppl 2 (1999) 3-7).

HPX is the major vehicle for the transportation of heme in the plasma and its principal role is to prevent heme-mediated oxidative stress and loss of heme-bound iron (Tolosano, E. & Altruda, F., DNA Cell Biol 21 (2002) 297-306). It can protect cells against oxidative stress by inducing the expression of intracellular antioxidants such as heme oxygenase, metallothioneins and ferritin. Metallothioneins are cytosolic proteins that are expressed particularly in synovial fibroblasts (Backman, J.T. et al., Virchows Arch 433 (1998) 153-160). There is significant experimental evidence for the presence of oxidative stress in the synovial tissue of RA patients (reviewed in: Schett, G. et al., Arthritis Res 3 (2000) 80-86). Furthermore HPX was reported to promote proliferation of human T lymphocytes (Smith, A. et al., Exp Cell Res 232 (1997) 246-254). These studies render it likely that HPX belongs to the up-regulated proteins in serum and synovia of RA patients, thereby providing a rationale for the relevance of HPX (351-367) as a RA-specific candidate marker.

Hsc70-interacting protein

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An epitope which was mostly identified in serum samples (4 out of 7 erosive and non-erosive RA samples) and which is also related to stress responses is derived from the Hsc70-interacting protein Hip: Hip (83-98) with the amino acid sequence of SEQ ID NO: 38 (Table 3). Two length variants of this epitope were identified (Table 3): the 18-mer Hip (83-100; SEQ ID NO: 36) and the 15-mer Hip (84-98; SEQ ID NO: 39). An additional

length variant was discovered in one erosive synovia sample (Table 3): the 15-mer Hip (85-99; SEQ ID NO: 37). As judged from the shortest length variant Hip (84-98) a DRB1*0401 binding motif attaining a moderate score of 8%, can be postulated: 89I as P1 anchor, 92D as P4 anchor, 94D as P6 anchor.

In the cytosol of eukaryotic cells, Hip and Hop proteins associate with Hsc70 in order to participate in the regulation of Hsc70 chaperone activity (Frydman, J. & Höhfeld, J., Trends Biochem Sci 22 (1997) 87-92). The 42 kD Hip protein binds to the ATPase domain of Hsc70. It was postulated that Hip might increase the half-life of the chaperone-substrate complex providing the molecular basis for an efficient cooperation of Hsc70 with downstream chaperone systems. Hsc70 and Hsp90 have been shown to cooperate during protein folding *in vitro* (Jakob, U. & Buchner, J., Trends Biochem Sci 19 (1994) 205-211; Freeman, B.C. & Morimoto, R.I., EMBO J 15 (1996) 2969-2979) and to play a role in thermal denaturation (Schneider, C. et al., PNAS 93 (1996) 14536-14541). The Hsc70 and Hsp90 association with stress-adaptation ultimately links Hip to stress responses, including the induction of heat shock proteins, in the synovial tissue of RA patients (reviewed in: Schett, G. et al., Arthritis Res 3 (2001) 80-86).

Table 1: HLA-DR associated peptide antigens from serum and synovial fluid of patients with mostly non-erosive RA.

Protein source ⁸	Interferon-gamma-inducible lysosomal thiol reductase (192-205)	Influenza Haemagglutinin (307-319)	Immunglobuline kappa (188-202)	M. tuberculosis Hsp65 (3-13)
DRB1*0401- binding score ^f	1%	1%	%7	> 10%
Sequence	GDRGMOEMHANAOR GDRGMOEMHANAORTD GDRGMOEMHANAORTD GDRGMOEMHANAORTD GDRGMOEMHANAORTD	PK <u>INKONTEKA</u> AT ^{h ()}	KHKV <u>YAĞEMIĞİQĞ</u> LS ^{h (ii)}	KTĪMKDBERARRI
Haplo- Length	14 17 16 17 16			
Haplo-	1 E E 4 4 E	la la	inder	
Sample	S Sym Sym	strong HLA-DRB1*0401 binder	moderate HLA-DRB1*0401 binder	weak HLA-DRB1*0401 binder
	6.8 6.8 9.1 9.1 20.7	HLA-DRB1	rate HLA-D)	HLA-DRB1'
RA-	Z Z Z Z Z E	strong	moder	weak I
SEQ. ID. RA-	NO.	,		

*RA-type of the patient based on clinical diagnosis: persistant erosive (E) or persistent non-erosive (N) RA

^bRheumatoid factor

Sample description: dendritic cells pulsed with serum (S) or synovial fluid (Syn)

^dHaplotype of the buffy coat: (1) HLA-DRB1*0401, *03011; (2) HLA-DRB1*0401, *0304; (3) HLA-DRB1*0401, *1301; (4) HLA-DRB1*0401, *0701

'Sequences of the RA-derived peptides in one-letter-code. The HLA-DRB1*0401 binding motif is boxed in grey.

Score of the epitope in context of the HLA-DRB1*0401 allele based on the TEPITOPE program (Hammer, J. et al., Adv Immunol 66 (1997) 67-100).

h (i) Rothbard, J.B. et al., Cell 52 (1988) 515-523. h (ii) Chicz, R.M. et al., J Exp Med 178 (1993) 27-47. h (iii) van Schooten, W.C. et al., Eur J Immunol 19 (1989) 2075-2079. ** Sprotein name according to the Swiss-Prot / TrEMBL database. The numbers in brackets represent the shortest length variant of the respective epitope. 2

Table 2: HLA-DR associated peptide antigens from serum and synovial fluid of patients with mostly erosive RA.

								•				
	DICBI*0401- Protein source		Amelian	Aponpoprotein 15-100 (2877-2892)			•					
	DICE 1*0401-	omaing score	%	2								
Haplo Length Consense	הלתכווכנ	•	INNOLTIED SNITKYFHK**		INNOETEDSNITKYFHK**	INNQUELIDSNITIKNEHK**		INNOLTED SNITKY PHK**	N. I.O. F. HIVE SECTION AND ADDRESS OF THE PARTY OF THE P	ININCELLUSINITERIEHKI.	INNOL'ELDSNITKYFHK**	A STATE OF THE PARTY OF THE PAR
Lenoth	119112		16	71	97	16		16		` `	16	
Haplo	-type ^d		-	Ç	1	3	•	m		•	1	
Sample			S	S	,	S	c	n	S		S	
RF ^b	(IU/ml)			+		134	202	7.07	20.7		•	
RA-	typeª		ធា	ய	1	ш	ÇT.	1	ല	;	z	
SEQ. ID.	NO.		4	4	•	4	4		2	•	4.	

*RA-type of the patient based on clinical diagnosis: persistant erosive (E) or persistent non-erosive (N) RA

^bRheumatoid factor

Sample description: dendritic cells pulsed with serum (S) or synovial fluid (Syn)

^dHaplotype of the buffy coat: (1) *HLA-DRB1*0401*, *03011; (2) *HLA-DRB1*0401*, *0304; (3) *HLA-DRB1*0401*, *1301; (4) *HLA-DRB1*0401*, *0701

*Sequences of the RA-derived peptides in one-letter-code. The HLA-DRB1*0401 binding motif is boxed in grey.

⁸Protein name according to the Swiss-Prot / TrEMBL database. The numbers in brackets represent the shortest length variant of the respective epitope. ¹Score of the epitope in context of the HLA-DRB1*0401 allele based on the TEPITOPE program (Hammer, J. et al., Adv Immunol 66 (1997) 67-100).

** Length variant of the respective epitope, which was identified in 2 healthy control samples as well.

fluid of patients with erosive and non-erosive RA. Table 3: HLA-DR associated peptide antigens from serum or synvial

Protein source ^s	Inter-alpha-trypsin inhibitor	heavy chain H4	(274-287)														Complement C4	(102/1-/601)			
DRB1*0401- binding score ^f	1%																1%				
Length Sequence	NATIONAL PROPERTY OF STATE OF	MPKN V VENTER SEASON	MPKNVVFVIDKSGSMSGK	MPKNVVFVIDKSGSMSG	MPKNVVEVIDKSGSMSG	NVVFVIDKSGSMSG	MPKNVVFVIDKSGSMSG	KNVVFVIDKSESMSG	MPKNVVEVIDKSGSMSG	NVVEVIDKSGSMSGR	MPKNVVEVIDKSGSMSGRK	MPKNVVFVIDKSGSMSGK	MPKNVVFVIDKSGSMSG	MPKNVVFVIDKSGSMSG	NVVFWIDKSGSMSGRK	MPKNVVFVIDKSGSMSG	GHPQXIEDSNSWIBE**	GHPQ <u>YELDSNSWI</u> BEM	GHPQYLIDSNSWIE	GHPOXLLDSNSWIEE**	
Length		I9	18	17	17	14	17	15	17	15	. 19	18	17	17	16	17	15	16	14	7.	3
Haplo-	type	m	<u>ო</u>	m	ю	က	7	2	1		en	ю	3	æ	ю	-		1		"	n
Sample		S	s	S	S	s	S	S	S	S	S	s	S	s	S	S	S	S	S		n
	(IU/ml)		+	+	134	. 134	134	134	20.7	20.7	•	8.9	6.8	8.9	8.9	8.9	•	•	•	,	6.8
RA-	type	ш	ш	ш	ш	ដោ	దు	щ	ជា	ш	z	Z	z	Z	Z	Z	z	Z	; 2	5	Z
SEQ. ID	No.	9	7	∞	∞	6	∞	10	oc	> =	۰ ب	7	. œ	· «	12	∞	ũ	3 7	. 1	CI	13

													Complement C2		(C##1-1C#1)				CEI2 domestic Line	guidun-dunain cric	Brutanine acid-rich-like	protein 3 (15-26)
										٠			%6						4%	8		
GHPQXILIDSNSWIJEEMPS*	HPOWEEDSNSWIE	GHPQXLIDDSNSWIEE**	GHPQM	GHPQ<u>YEHDSNSWI</u>EEM	GHPQXIEDSNSWMEE**	GHPQNIEDSNSW	GHPQ <u>MIEDSWSWI</u> EE**	GHPQKEEDSINSWIEE**	GHPQYTHDSNSWIE	GHPQXIIIDSNSWIEEMPS*	GHPQNIEDSNSWIEE	GHPQ <u>XLIMBSNSWN</u> BEMPS*	GVDRYISK <u>YELDKAFSB</u> RN	RYISKAKEKDIKAKISDIR	ISKATELDIKATEDIN	ISKWEEDKAESDRNT	ISKAELDKARSDR	ISK <u>veldkafsb</u> rn	GSREJKS@@SEVARRILY	REIKS O SEVETIR	GSREITKS@@SEVARR	GSREIKSQQSENTINT*
18	13	15	12	16	15	12	15	15	14	18	15	18	19	15	14	15	13	14	16	12	14	16
4	4	4	4		1		2	8	9	m	8	8	4	7	-	-	7	7	က	7	-	-
S	တ	S	S	S	S	S	S	S	S	တ	S	S	S	တ	S	S	S	S	S	S	S	S
9.1	9.1	9.1	9.1	•	•	•	+	134	134	134	20.7	20.7	9.1	•	•	•	ı	+	8.9	•	•	,
z	z	z	z	ш	ធា	ш	щ	ш	ш	ш	ш	ш	Z	Z	Z	Z	Z	បា	z	z	Z	z
16	12	13	8 2	<u> </u>	E	8 2	13	13	15	16	13	16	19	70	21		23	21	24	25	5 6	24

	Interleukin-4- induced protein 1 (293-308)	Hemopexin (351-363)
	1%	1%
REFKSQQSEWIPUL GSREIKSQQSEWIP REFKSQQSEWIP REFKSQQSEWIP REFKSQQSEWIP GSREFKSQQSEWIP	GPHDVHWQIEISPPARNIK GPHDVHWQIEISPPARNIK GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN GPHDVHWQIEISPPARN	GPHDVHWQLEFSFERM GPHDVHWQLETSBPARN TPHGILIDSWDAMFICPG TPHGILIDSWDAMFICP** TPHGILIDSWDAMFICP**
14 14 12 12 12	17 19 17 17 16 16 17	17 17 17 18 17
		1 4 6 1 4 4
ν ν ν ν ν	Syn Syn S S S S S S S S S S S S S S S S	S Syn S
- - 134 20.7 20.7	20.7 20.7 134 + 134 134 20.7 6.8	9.1 6.8 - 9.1
Z 11 12 12 12 12 12	ы ы ы ы ы ы ы и и и и и и и и и и и и и	z z z z z z
27 26 25 25 25 24	30 30 30 30 30 30 30 30 30 30 30 30 30 3	28 28 28 31 32 33

												Hsc70-interacting protein	(84-98)					
				•							į	%8						
TPHGWWWWW	TPHCIMING WATER TOTAL	TPHGHINDSWA	CTPHCETT SOVEMENT	TPH GITTED SYNDAMBER*	TPH CHIED CARTESTON**	TPHGIMIDAVIDATEICE	TPHGIII PSVIDAMEI*	TPHCATTANCVER CONT.	TPHGIRBOXTOATER	TPHGIIIFDSWIDAMEI CD**	IDKEGVI用PDARBARDAR	KECVIBBRIDAND	IDKFCV/IEDE/EDIA		DAEGVER STREET S	TOVE CVITTE THE TOWN OF THE PROPERTY OF THE PR	IDAGO NIEKO MARKETE	1 D N D C V II THE TO SEE SEE SEE
. 17	17	13	14	15	17	81	15	17	: 12	17	18	15	16	<u> </u>	3 %	<u> </u>	<u>י</u>	>
ю	4	4	4	ო	m	7	m	က	m	m	m	<u>ო</u>	-	-	• 4	٠ -	· "	
Syn	Syn	Syn	Syn	S	S	S	S	S	S	S	Syn	Syn	တ	S	S	S	S	
8.9	9.1	9.1	9.1	8.9	6.8	+	134	134	20.7	20.7	134	134	•	•	9.1	,	8.9	
											ш							
32	32	33	34	35	32	31	35	32	35	32	36	37	38	39	36	38	38	

RA-type of the patient based on clinical diagnosis: persistant erosive (E) or persistent non-erosive (N) RA

Sequences of the RA-derived peptides in one-letter-code. The HLA-DRBI*0401 binding motif is boxed in grey.
Score of the epitope in context of the HLA-DRBI*0401 allele based on the TEPITOPE program (Hammer, J. et al., Adv Immunol 66 (1997) 67-100).
Protein name according to the Swiss-Prot / TrEMBL database. The numbers in brackets represent the shortest length variant of the respective epitope. Sample description: dendritic cells pulsed with serum (S) or synovial fluid (Syn)

"Haplotype of the buffy coat: (1) HLA-DRB1*0401, *03011; (2) HLA-DRB1*0401, *0304; (3) HLA-DRB1*0401, *1301; (4) HLA-DRB1*0401, *0701

* (**) Length variant of the respective epitope which was identified in 1 (2) healthy control sample(s) as well.

y of the candidate RA markers. Table 4: Su

	t in the Accession number ples ^b	N P13284		6 E P04114	2 of 5 N Q14624	3 of 5 N P01028	2 of 5 N P01024	3 of 6 E / 2 of 5 N Q9H299	6 of 6 E / 5 of 5 N Q96RQ9	3 of 6 E / 5 of 5 N P02790	2 of 6 E / 3 of 5 N P50502
	Frequency in the RA samples ^b	3 of 5 N		4 of 6 E	4 of 6 E / 2 of 5 N	4 of 6 E / 3 of 5 N	1 of 6 E / 2 of 5 N	3 of 6 E /	6 of 6 E	3 of 6 E	2 of 6 E
ndidate KA iliaincis.	Protein source	doctor Late 1	Interferon-gamma-inducible lysosomal thiol reductase	Apolipoprotein B-100	Inter-alpha-trypsin inhibitor heavy chain H4	Complement C4	Complement C3	SH3 domain-binding glutamic acid-rich-like protein 3	Interleukin-4-induced protein 1	Hemopexin	II.270 interacting protein
able 4: Summary of the candidate ICA man	RA-type		mostly non-erosive	mostly prosive	dhistra was pro-	erosive and non-crosive					

*Protein name according to the Swiss-Prot / TrEMBL database.

^bFrequency of the identified epitope in the RA samples. The RA-type of the patient was based on clinical diagnosis: persistant erosive (E) or persistent non-erosive (N) RA.

relates to the Swiss-Prot database. S

Claims

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- 1. A MHC class II antigenic peptide comprising
 - (a) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57, or
 - (b) at least the amino acid sequence of the peptide binding motif selected from the group consisting of SEQ ID NOs. 49 to 57 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 39.
- 2. A MHC class II antigenic peptide comprising
- (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO.49, or
 - (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 49 with additional N-and C-terminal flanking sequences of a corresponding sequence selected from the group consisting of SEQ ID NOs. 1 to 3.
- 15 3. A MHC class II antigenic peptide comprising
 - (a) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50, or
 - (b) at least the amino acid sequence of the peptide binding motif of SEQ ID NO. 50 with additional N-and C-terminal flanking sequences of the corresponding sequence of SEQ ID NO. 5.
 - 4. The MHC class II antigenic peptide according to any one of claims 1 to 3 linked to a MHC class II molecule.
 - 5. An antibody reactive with a MHC class II antigenic peptide according to any one of claims 1 to 3.
- 25 6. A nucleic acid molecule encoding a peptide or polypeptide according to any one of claims 1 to 4.
 - 7. A recombinant nucleic acid construct comprising the nucleic acid molecule according to claim 6 operably linked to an expression vector.
 - 8. A host cell containing the nucleic acid construct according to claim 7.
- 9. A method for producing a MHC class II antigenic peptide according to any one of claims 1 to 3 comprising the steps of culturing the host cell of claim 8 under

conditions allowing expression of peptide from the cells or the culture

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said peptide and recovering the medium.

- 10. A method for isolating and identifying MHC class II associated RA antigenic peptides in femtomolar amounts, which method comprises
 - (a) providing immature dendritic cells in a number comprising 0.1 to 5 µg MHC class II molecules;
 - (b) contacting the cells of (a) with serum or synovial fluid and inducing maturation of dendritic cells by adding TNFalpha;
- (c) isolating class II MHC molecule-antigenic peptide complexes from the cells with methods comprising solubilization of the cells and sequestration of the complexes of MHC class II molecules with antigenic peptides by immunoprecipitation or immunoaffinity chromatography;
 - (d) washing the sequestered complexes of MHC class II molecules with antigenic peptides with water in an ultrafiltration tube;
 - (e) eluting the associated antigenic peptides from the MHC class II molecules at 37°C with diluted trifluoro acetic acid, and
 - (f) separating, detecting and identifying the isolated peptides by liquid chromatography and mass spectrometry.
- 11. The method according to claim 10, wherein in step (f) of the method the liquid chromatography comprises a first linear elution step from the reversed-phase material with a volume sufficient to elute contaminants prior to the peptide elution step.
- 12. The method according to any one of claims 10 and 11, further comprising (g) analyzing the identified peptides by methods comprising a database and a software developed to perform comparative data analysis across multiple datasets.
- 13. A pharmaceutical composition comprising a MHC class II antigenic peptide according to any one of claims 1 to 3, an antibody according to claim 5, or a polypeptide selected from the group consisting of SEQ ID NOs 40 to 48, and optionally a pharmaceutically acceptable carrier.
- 30 14. A diagnostic composition comprising the antibody according to claim 5.
 - 15. The use of the MHC class II antigenic peptide according to claim 1, wherein the antigenic peptide is a marker for erosive and/or non-erosive RA.
 - 16. The use of the MHC class II antigenic peptide according to claim 2, wherein the antigenic peptide is a marker for non-erosive RA

17. The use of the MHC class II wherein the antigenic peptide is a

antigenic peptide according claim 3, marker for erosive RA.

- 18. The use of a polypeptide selected from the group consisting of SEQ ID NOs 40 to 48 as a marker for RA, preferably for erosive and/or non-erosive RA.
- 19. The antigenic peptides, antibodies, nucleic acids, host cells, methods, compositions and uses substantially as herein before described especially with reference to the foregoing Examples.

Abstract

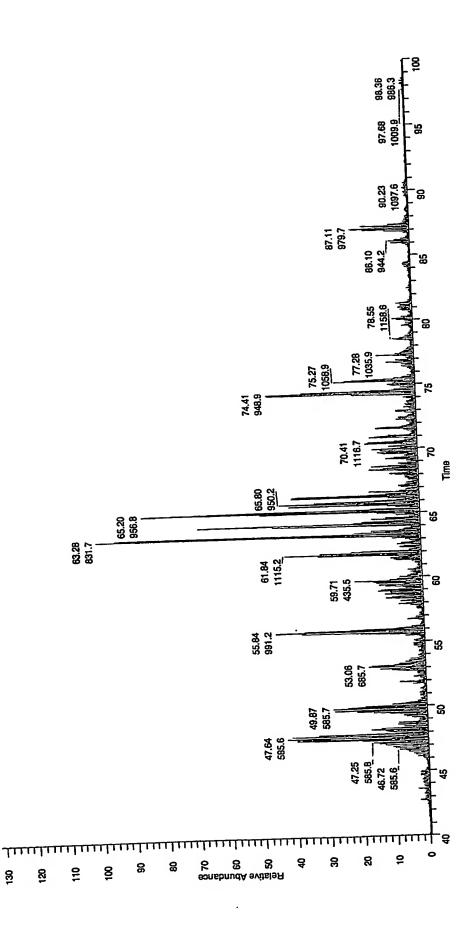
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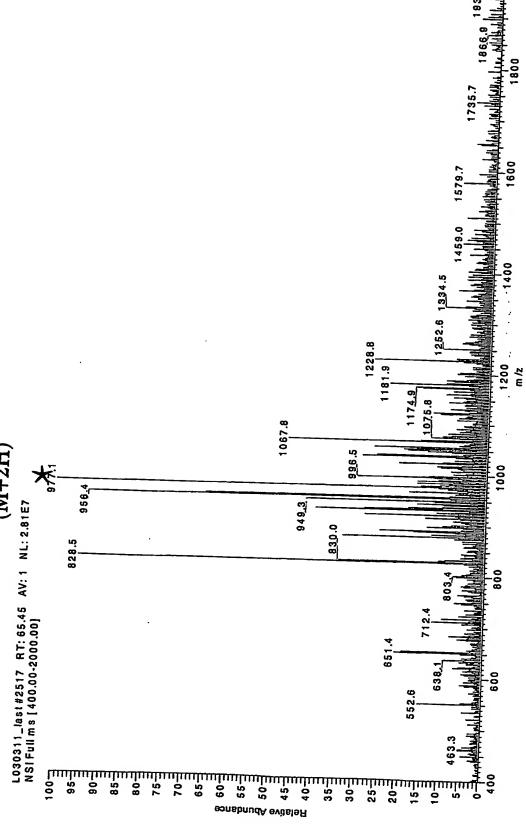
The present invention provides novel naturally-processed MHC class II antigenic peptides which originate from interferon-γ-inducible lysosomal thiol reductase, apolipoprotein B-100, inter-α-trypsin inhibitor heavy chain H4, complement C4, complement C3, SH3 domain-binding glutamic acid-rich-like protein 3, interleukin-4-induced protein 1, hemopexin, and Hsc70-interacting protein. Also provided are these antigenic peptides and the proteins they are derived from as markers for erosive and/or non-erosive RA. Moreover, these antigenic peptides linked to MHC class II molecules, antibodies reactive with said antigenic peptides, nucleic acids encoding said antigenic peptides, and nucleic acid constructs, host cells and methods for expressing said antigenic peptides are provided. The antigenic peptides of the invention can be used as markers in diagnosis of RA and in therapy as anti-RA vaccines.

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Fig.]

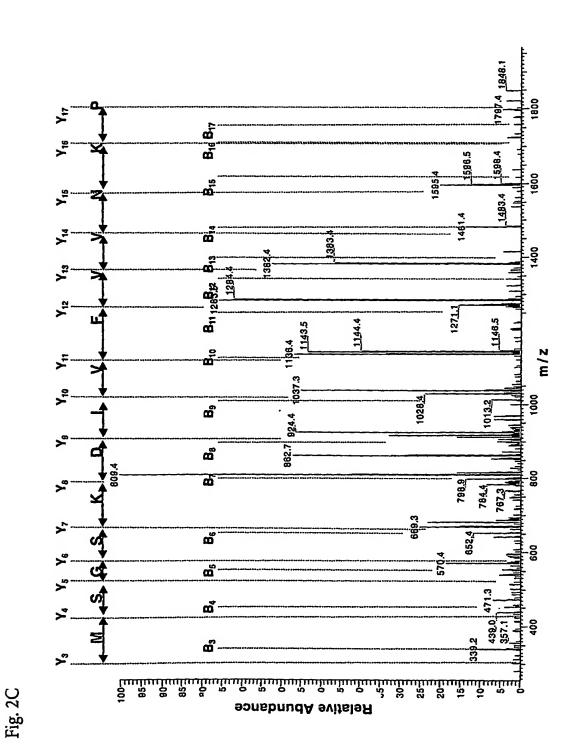
Fig. 2A





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Fig. 2B



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Asn Arg Gln Ser Met Thr Leu Ser Ser 1055 Glu Val Gln Ile Pro Asp 1065 Phe Asp Val Asp Leu Gly Thr Ile Leu Arg Val Asn Asp Glu Ser 1070 1080 Thr Glu Gly Lys Thr Ser Tyr Arg Leu Thr Leu Asp Ile Gln Asn 1085 1090 1095 10 Lys Lys Ile Thr Glu Val Ala Leu Met Gly His Leu Ser Cys Asp 1100 1110 15 Thr Lys Glu Glu Arg Lys Ile Lys Gly Val Ile Ser Ile Pro Arg 20 Leu Gln Ala Glu Ala Arg Ser Glu Ile Leu Ala His Trp Ser Pro 1130 1135 1140 25 Ala Lys Leu Leu Gln Met Asp Ser Ser Ala Thr Ala Tyr Gly 1145 1150 Ser Thr Val Ser Lys Arg Val Ala Trp His Tyr Asp Glu Glu Lys 1160 1165 30 Ile Glu Phe Glu Trp Asn Thr Gly Thr Asn Val Asp Thr Lys Lys 1175 1180 1185 35 Met Thr Ser Asn Phe Pro Val Asp Leu Ser Asp Tyr Pro Lys Ser 1190 1195 1200 40 Leu His Met Tyr Ala Asn Arg Leu Leu Asp His Arg Val Pro Glu 1205 1210 45 Thr Asp Met Thr Phe Arg His Val Gly Ser Lys Leu Ile Val Ala Met Ser Ser Trp Leu Gln Lys Ala Ser Gly Ser Leu Pro Tyr Thr 1235 1240 1245 50 Gln Thr Leu Gln Asp His Leu Asn Ser Leu Lys Glu Phe Asn Leu 1250 . 1255 1250 . 55 Gln Asn Met Gly Leu Pro Asp Phe His Ile Pro Glu Asn Leu Phe 60 Leu Lys Ser Asp Gly Arg Val Lys Tyr Thr Leu Asn Lys Asn Ser 1280 1285 65 Leu Lys Ile Glu Ile Pro Leu Pro Phe Gly Gly Lys Ser Ser Arg 1300 Asp Leu Lys Met Leu Glu Thr Val Arg Thr Pro Ala Leu His Phe 70 Lys Ser Val Gly Phe His Leu Pro Ser Arg Glu Phe Gln Val Pro 1325 1330 1335 1325 75

Thr Phe Thr Ile Pro Lys Leu Tyr Gln Leu Gln Val Pro Leu Leu

5	Gly Val Leu Asp Leu Ser Thr Asn Val Tyr Ser Asn Leu Tyr Asn 1355 1360 1365
	Trp Ser Ala Ser Tyr Ser Gly Gly Asn Thr Ser Thr Asp His Phe 1370 1375 1380
15	
	Leu Leu Ser Tyr Asn Val Gln Gly Ser Gly Glu Thr Thr Tyr Asp 1400 1405 1410
20	His Lys Asn Thr Phe Thr Leu Ser Cys Asp Gly Ser Leu Arg His 1415 1420 1425
25	Lys Phe Leu Asp Ser Asn Ile Lys Phe Ser His Val Glu Lys Leu 1430 1440
30	Gly Asn Asn Pro Val Ser Lys Gly Leu Leu Ile Phe Asp Ala Ser 1445 1450 1455
35	Ser Ser Trp Gly Pro Gln Met Ser Ala Ser Val His Leu Asp Ser 1460 1465 1470
<i>JJ</i>	Lys Lys Cln His Leu Phe Val Lys Glu Val Lys Ile Asp Gly 1475 1480 1485
40	Gln Phe Arg Val Ser Ser Phe Tyr Ala Lys Gly Thr Tyr Gly Leu 1490 1495 1500
45	Ser Cys Gln Arg Asp Pro Asn Thr Gly Arg Leu Asn Gly Glu Ser 1505 1510 1515
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55	Thr Gly Arg Tyr Glu Asp Gly Thr Leu Ser Leu Thr Ser Thr Ser 1535 1540 1545
<i>55</i>	Asp Leu Gln Ser Gly Ile Ile Lys Asn Thr Ala Ser Leu Lys Tyr 1550 1560
60	Glu Asn Tyr Glu Leu Thr Leu Lys Ser Asp Thr Asn Gly Lys Tyr 1565 1570 1575
65 [°]	Lys Asn Phe Ala Thr Ser Asn Lys Met Asp Met Thr Phe Ser Lys 1580 1585
70	Gln Asn Ala Leu Leu Arg Ser Glu Tyr Gln Ala Asp Tyr Glu Ser 1595 1600 1605
	Leu Arg Phe Phe Ser Leu Leu Ser Gly Ser Leu Asn Ser His Gly 1610
75	Leu Glu Leu Asn Ala Asp Ile Leu Gly Thr Asp Lys Ile Asn Ser 1625 1630 1635

5	GIÀ	1640	HIS	Lys	Ala	Thr	1645	Arg	11e		GTĀ	1650	.sp	GIĀ	TIE	56
	Thr	Ser 1655	Ala	Thr	Thr	Asn	Leu 1660	Lys	Cys	Ser	Leu	Leu 1665	Val	Leu	Glu	
10	Asn	Glu 1670	Leu	Asn	Ala	Glu	Leu 1675	Gly	Leu	Ser	Gly	Ala 1680	Ser	Met	Lys	
15	Leu	Thr 1685	Thr	Asn	Gly	Arg	Phe 1690	Arg	Glu	His	Asn	Ala 1695	Lys	Phe	Ser	
20	Leu	Asp 1700	Gly	Lys	Ala	Άla	Leu 1705	Thr	Glu	Leu	Ser	Leu 1710	Gly	Ser	Ala	
25	Tyr	Gln 1715	Ala	Met	lle	Leu	Gly 1720	Val	Asp	Ser	Lys	Asn 1725	Ile	Phe	Asn	
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4 0	Tyr	Ser 1775		Asp	Lys	Phe	Tyr 1780	Lys	Gln	Thr	Val	Asn 1785	Leu	Gln	Leu	
45	Gjn	Pro 1790		Ser	Leu	Val	Thr 1795		Leu	Asn	Ser	Asp 1800	Leu	Lys	Tyr	
	Asn	Ala 1805		Asp	Leu	Thr	Asn 1810	Asn	Gly	Lys	Leu	Arg 1815		Glu	Pro	I
50	Leu	Lys 1820		His	Val	Ala	Gly 1825		Leu	Lys	Gly	Ala 1830		Gln	Asn	l
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60	Ala	Ser 1850		Lys	Ala	Asp	Thr 1855		Ala	Lys	Val	Gln 1860		Val	Glu	l
65	Phe	Ser 1865		Arg	Leu	Asn	Thr 1870		Ile	Ala	Gly	Leu 1875		Ser	Ala	L
	Ile	Asp 1880		Ser	Thr	Asn	Tyr 1885		Ser	Asp	Ser	Leu 1890		Phe	Ser	•
70	Asn	Val 1895		Arg	Ser	Val	Met 1900		Pro	Phe	Thr	Met 1905		: Ile	Asp	•
75	Ala	His 1910		Asn	Gly	Asn	Gly 1915		Leu	Ala	Leu	Trp 1920		/ Glu	His	•

Thr Gly Gln Leu Tyr Ser Lys Phe Leu Leu Lys Ala Glu Pro Leu 1925 1930 1935 5 Ala Phe Thr Phe Ser His Asp Tyr Lys Gly Ser Thr Ser His His 1940 1945 1950 Leu Val Ser Arg Lys Ser Ile Ser Ala Ala Leu Glu His Lys Val 10 1960 Ser Ala Leu Leu Thr Pro Ala Glu Gln Thr Gly Thr Trp Lys Leu 1970 1980 15 Lys Thr Gln Phe Asn Asn Asn Glu Tyr Ser Gln Asp Leu Asp Ala 1985 1990 1995 20 Tyr Asn Thr Lys Asp Lys Ile Gly Val Glu Leu Thr Gly Arg Thr 2000 2005 2010 25 Leu Ala Asp Leu Thr Leu Leu Asp Ser Pro Ile Lys Val Pro Leu 2015 30 Leu Leu Ser Glu Pro Ile Asn Ile Ile Asp Ala Leu Glu Met Arg 2035 Asp Ala Val Glu Lys Pro Gln Glu Phe Thr Ile Val Ala Phe Val 2045 2055 35 Lys Tyr Asp Lys Asn Gln Asp Val His Ser Ile Asn Leu Pro Phe 2060 2070 40 Phe Glu Thr Leu Gln Glu Tyr Phe Glu Arg Asn Arg Gln Thr Ile 2075 2080 2085 2080 45 Ile Val Val Glu Asn Val Gln Arg Asn Leu Lys His Ile Asn 2090 2095 2100 50 Ile Asp Gln Phe Val Arg Lys Tyr Arg Ala Ala Leu Gly Lys Leu 2105 Pro Gln Gln Ala Asn Asp Tyr Leu Asn Ser Phe Asn Trp Glu Arg 2120 2130 55 Gln Val Ser His Ala Lys Glu Lys Leu Thr Ala Leu Thr Lys Lys 60 Tyr Arg Ile Thr Glu Asn Asp Ile Gln Ile Ala Leu Asp Asp Ala 2150 2155 2160 65 Lys Ile Asn Phe Asn Glu Lys Leu Ser Gln Leu Gln Thr Tyr Met 2165 2170 2175 70 Ile Gln Phe Asp Gln Tyr Ile Lys Asp Ser Tyr Asp Leu His Asp 2180 2185 2190 Leu Lys Ile Ala Ile Ala Asn Ile Ile Asp Glu Ile Ile Glu Lys 2195 2200 2205 75

Leu Lys Ser Leu Asp Glu His Tyr His 2210 2215 Ile Arg Val Asn Leu Val 5 Lys Thr Ile His Asp Leu His Leu Phe Ile Glu Asn Ile Asp Phe Asn Lys Ser Gly Ser Ser Thr Ala Ser Trp Ile Gln Asn Val Asp 2240 2245 2250 10 Thr Lys Tyr Gln Ile Arg Ile Gln Ile Gln Glu Lys Leu Gln Gln 2255 2260 2265 15 Leu Lys Arg His Ile Gln Asn Ile Asp Ile Gln His Leu Ala Gly 2270 2280 20 Lys Leu Lys Glm His Ile Glu Ala Ile Asp Val Arg Val Leu Leu 2290 25 Asp Gln Leu Gly Thr Thr Ile Ser Phe Glu Arg Ile Asn Asp Val 2300 2305 2310 Leu Glu His Val Lys His Phe Val Ile Asn Leu Ile Gly Asp Phe 2315 2320 2325 30 Glu Val Ala Glu Lys Ile Asn Ala Phe Arg Ala Lys Val His Glu 2330 2335 2340 35 Leu Ile Glu Arg Tyr Glu Val Asp Gln Gln Ile Gln Val Leu Met 2345 2350 2355 40 Asp Lys Leu Val Glu Leu Thr His Gln Tyr Lys Leu Lys Glu Thr 2360 2365 . 2370 Ile Gln Lys Leu Ser Asn Val Leu Gln Gln Val Lys Ile Lys Asp 2375 2380 2385 45 Tyr Phe Glu Lys Leu Val Gly Phe Ile Asp Asp Ala Val Lys Lys 2390 2395 50 Leu Asn Glu Leu Ser Phe Lys Thr Phe Ile Glu Asp Val Asn Lys 2410 2405 55 Phe Leu Asp Met Leu Ile Lys Lys Leu Lys Ser Phe Asp Tyr His 2430 60 Gln Phe Val Asp Glu Thr Asn Asp Lys Ile Arg Glu Val Thr Gln 2435 2445 65 Arg Leu Asn Gly Glu Ile Gln Ala Leu Glu Leu Pro Gln Lys Ala Glu Ala Leu Lys Leu Phe Leu Glu Glu Thr Lys Ala Thr Val Ala 70 2470 Val Tyr Leu Glu Ser Leu Gln Asp Thr Lys Ile Thr Leu Ile Ile 2485 2480 75 Asn Trp Leu Gln Glu Ala Leu Ser Ser Ala Ser Leu Ala His Met

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5	Lys	Ala 2510	Lys)	Phe	Arg	Glu	Thr 2515	Leu	Glu	Asp	Thr	Arg 2520	Asp	Arg	Me't
10	Tyr	Gln 2525	Met	Asp	Ile	Gln	Gln 2530	Glu	Leu	Gln	Arg	Tyr 2535	Leu	Ser	Leu
15	Val	Gly 2540	Gln	Val	Tyr	Ser	Thr 2545	Leu	Val	Thr	Tyr	Ile 2550	Ser	Asp	Trp
15	Trp	Thr 2555	Leu	Ala	Ala	Lys	Asn 2560	Leu	Thr	Asp	Phe	Ala 2565		Gln	Tyr
20	Ser	Ile 2570	Gln	Asp	Trp	Ala	Lys 2575	Arg	Met	Lys	Ala	Leu 2580		Glu	Gln
25	Gly	Phe 2585	Thr	Val	Pro	Glu	Ile 2590	Lys	Thr	Ile	Leu	Gly 2595	Thr	Met	Pro
30	Ala	Phe 2600	Glu	Val	Ser	Leu	Gln 2605	Ala	Leu	Gln	Lys	Ala 2610	Thr	Phe	Gln
35	Thr	Pro 2615	Asp	Phe	Ile	Vaļ	Pro 2620	Leu	Thr	Asp	Leu	Arg 2625	Ile	Pro	Ser
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40	: Arg	Phe 2645	Ser	Thr	Pro	Glu	Phe 2650	Thr	Ile	Leu	Asn	Thr 2655	Phe	His	Ile
45	Pro	Ser 2660	Phe	Thr	Ile	Asp	Phe 2665	Val	Glu	Met	Lys	Val 2670	Lys	Ile	Ile
50	Arg	Thr 2675	Ile	Asp	Gln	Met	Gln 2680	Asn	Ser	Glu	Leu	Gln 2685	Trp	Pro	Val
55	Pro	Asp 2690	Ile	Tyr	Leu	Arg	Asp 2695	Leu	Lys	Val	Glu	Asp 2700	Ile	Pro	Leu
33	Ala	Arg 2705	Ile	Thr	Leu	Pro	Asp 2710	Phe	Arg	Leu	Pro	Glu 2715	Ile	Ala	Ile
60	Pro	Glu 2720	Phe	Ile	Ile	Pro	Thr 2725	Leu	Asn	Leu	Asn	Asp 2730	Phe	Gln	Val
65	Pro	Asp 2735	Leu	His	Ile	Pro	Glu 2740	Phe	Gln	Leu	Pro	His 2745	Ile	Ser	His
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75	Ile	Gln 2765	Ser	Pro	Leu	Phe	Thr 2770	Leu	Asp	Ala		Ala 2775	Asp	Ile	Gly
, 3	Asn	Gly 2780	Thr	Thr	Ser .	Ala	Asn 2785	Glu	Ala	Gly		Ala 2790	Ala :	Ser	Ile

Thr Ala Lys Gly Glu Ser Lys Leu Glu Val Leu Asn Phe Asp Phe 5 Gin Ala Asn Ala Gin Leu Ser Asn Pro Lys Ile Asn Pro Leu Ala 2810 2820 10 Leu Lys Glu Ser Val Lys Phe Ser Ser Lys Tyr Leu Arg Thr Glu 2835 2830 His Gly Ser Glu Met Leu Phe Phe Gly Asn Ala Ile Glu Gly Lys 15 Ser Asn Thr Val Ala Ser Leu His Thr Glu Lys Asn Thr Leu Glu 20 2855 Leu Ser Asn Gly Val Ile Val Lys Ile Asn Asn Gln Leu Thr Leu 2870 2880 25 Asp Ser Asn Thr Lys Tyr Phe His Lys Leu Asn Ile Pro Lys Leu 2895 Asp Phe Ser Ser Gln Ala Asp Leu Arg Asn Glu Ile Lys Thr Leu 2900 2905 2910 30 Leu Lys Ala Gly His Ile Ala Trp Thr Ser Ser Gly Lys Gly Ser 35 Trp Lys Trp Ala Cys Pro Arg Phe Ser Asp Glu Gly Thr His Glu 2930 2935 40 Ser Gln Ile Ser Phe Thr Ile Glu Gly Pro Leu Thr Ser Phe Gly 2945 2950 2955 45 Leu Ser Asn Lys Ile Asn Ser Lys His Leu Arg Val Asn Gln Asn 50 Leu Val Tyr Glu Ser Gly Ser Leu Asn Phe Ser Lys Leu Glu Ile 2975 2980 2985 Gln Ser Gln Val Asp Ser Gln His Val Gly His Ser Val Leu Thr 55 Ala Lys Gly Met Ala Leu Phe Gly Glu Gly Lys Ala Glu Phe Thr 3005 3010 3005 60 Gly Arg His Asp Ala His Leu Asn Gly Lys Val Ile Gly Thr Leu 3020 3025 3030 65 Lys Asn Ser Leu Phe Phe Ser Ala Gln Pro Phe Glu Ile Thr Ala 3035 3040 3045 Ser Thr Asn Asn Glu Gly Asn Leu Lys Val Arg Phe Pro Leu Arg 3050 3060 70 Leu Thr Gly Lys Ile Asp Phe Leu Asn Asn Tyr Ala Leu Phe Leu 75

Ser Pro Ser Ala Gln Gln Ala Ser Trp Gln Val Ser Ala Arg Phe 3080 3085 3090 5 Asn Gln Tyr Lys Tyr Asn Gln Asn Phe Ser Ala Gly Asn Asn Glu 3095 3100 3105 Asn Ile Met Glu Ala His Val Gly Ile Asn Gly Glu Ala Asn Leu 3110 3115 3120 10 Asp Phe Leu Asn Ile Pro Leu Thr Ile Pro Glu Met Arg Leu Pro 3125 15 Tyr Thr Ile Ile Thr Thr Pro Pro Leu Lys Asp Phe Ser Leu Trp 3140 3145 3150 20 Glu Lys Thr Gly Leu Lys Glu Phe Leu Lys Thr Thr Lys Gln Ser 25 Phe Asp Leu Ser Val Lys Ala Gln Tyr Lys Lys Asn Lys His Arg 3170 3175 3180 His Ser Ile Thr Asn Pro Leu Ala Val Leu Cys Glu Phe Ile Ser 30 3190 Gln Ser Ile Lys Ser Phe Asp Arg His Phe Glu Lys Asn Arg Asn 3200 3205 3210 35 Asn Ala Leu Asp Phe Val Thr Lys Ser Tyr Asn Glu Thr Lys Ile 3215 3220 3225 40 Lys Phe Asp Lys Tyr Lys Ala Glu Lys Ser His Asp Glu Leu Pro 3230 3230 Glu Leu Pro 45 Arg Thr Phe Gln Ile Pro Gly Tyr Thr Val.Pro Val Val Asn Val 3245 50 Glu Val Ser Pro Phe Thr Ile Glu Met Ser Ala Phe Gly Tyr Val 3260 3265 3270 Phe Pro Lys Ala Val Ser Met Pro Ser Phe Ser Ile Leu Gly Ser 3275 55 Asp Val Arg Val Pro Ser Tyr Thr Leu Ile Leu Pro Ser Leu Glu 3290 3295 3300 60 Leu Pro Val Leu His Val Pro Arg Asn Leu Lys Leu Ser Leu Pro 65 His Phe Lys Glu Leu Cys Thr Ile Ser His Ile Phe Ile Pro Ala 3320 3330 70 Met Gly Asn Ile Thr Tyr Asp Phe Ser Phe Lys Ser Ser Val Ile Thr Leu Asn Thr Asn Ala Glu Leu Phe Asn Gln Ser Asp Ile Val 75 3350

- Ala His Leu Leu Ser Ser Ser Ser Ser Val Ile Asp Ala Leu Gln 3365 3370 3375
- 5 Tyr Lys Leu Glu Gly Thr Thr Arg Leu Thr Arg Lys Arg Gly Leu 3380 3385 3390
- Lys Leu Ala Thr Ala Leu Ser Leu Ser Asn Lys Phe Val Glu Gly 3395 3400 3405
- Ser His Asn Ser Thr Val Ser Leu Thr Thr Lys Asn Met Glu Val 3410 3415 3420
 - Ser Val Ala Lys Thr Thr Lys Ala Glu Ile Pro Ile Leu Arg Met 3425 3430 3435
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 Asn Phe Lys Gln Glu Leu Asn Gly Asn Thr Lys Ser Lys Pro Thr 3440
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- Leu Tyr Ser Thr Ala Lys Gly Ala Val Asp His Lys Leu Ser Leu 30 3470 3480
- Glu Ser Leu Thr Ser Tyr Phe Ser Ile Glu Ser Ser Thr Lys Gly 3485 3490 3495 .
 - Asp Val Lys Gly Ser Val Leu Ser Arg Glu Tyr Ser Gly Thr Ile 3500 3505
- 40
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 3515 3520 3525
- 45 Ser Val Lys Leu Gln Gly Thr Ser Lys Ile Asp Asp Ile Trp Asn 3530 3540
- Leu Glu Val Lys Glu Asn Phe Ala Gly Glu Ala Thr Leu Gln Arg 3545 3550 3555
- Ile Tyr Ser Leu Trp Glu His Ser Thr Lys Asn His Leu Gln Leu 3560 3565 3570
 - Glu Gly Leu Phe Phe Thr Asn Gly Glu His Thr Ser Lys Ala Thr 3575 3580 3585
- 60 Leu Glu Leu Ser Pro Trp Gln Met Ser Ala Leu Val Gln Val His 3590 3600
- 65 Ala Ser Gln Pro Ser Ser Phe His Asp Phe Pro Asp Leu Gly Gln 3605 3610 3615
- Glu Val Ala Leu Asn Ala Asn Thr Lys Asn Gln Lys Ile Arg Trp 3620 3625 3630

- Lys Asn Glu Val Arg Ile His Ser Gly Ser Phe Gln Ser Gln Val 3635 3640 3645
 - Glu Leu Ser Asn Asp Gln Glu Lys Ala His Leu Asp Ile Ala Gly

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10	Val	Tyr 3680	Asp	Lys	Ser	Leu	Trp 3685	Asp	Phe	Leu	Lys	Leu 3690		Val	Thr
15	Thr	Ser 3695	Ile	Gly	Arg	Arg	Gln 3700	His	Leu	Arg	Val	Ser 3705	Thr	Ala	Phe
15	Va1	Tyr 3710	Thr	Lys	Asn	Pro	Asn 3715	Gly	Tyr	Ser	Phe	Ser 3720	Ile	Pro	Val
20	Lys	Val 3725	Leu	Ala	Asp	Lys	Phe 3730	Ile	Thr	Pro	Gly	Leu 3735	Lys	Leu	Asn
25	Asp	Leu 3740	Asn	Ser	Val	Leu	Val 3745	Met	Pro	Thr	Phe	His 3750	Val	Pro	Phe
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35	Gln	Ile 3770	Tyr	Lys	Lys	Leu	Arg 3775	Thr	Ser	Ser	Phe	Ala 3780	Leu	Asn	Leu
<i>JJ</i>	Pro	Thr 3785	Leu	Pro	Glu	Val	Lys 3790	Phe		Glu	Val	Asp 3795	Val	Leu	Thr
40	Lys	туr 3800	Ser	Gln	Pro	Glu	Asp 3805	Ser	Leu	Ile	Pro	Phe 3810	Phe	Glu	Ile
45	Thr	Val 3815	Pro	Glu	Ser	Gln	Leu 3820	Thr	Val	Ser	Gln	Phe 3825	Thr	Leu	Pro
50	Lys	Ser 3830	Val	Ser	Asp	Gly	Ile 3835	Ala	Ala	Leu	Asp	Leu 3840	Asn	Ala	Val
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7 5	Ser	Thr 3920	Val	Gln	Phe	Leu	Glu 3925	Tyr	Glu	Leu	Asn	Val 3930	Leu	Gly	Thr
, ,	His	Lys 3935	Ile	Glu	Asp	Gly	Thr 3940	Leu	Ala	Ser	Lys	Thr 3945	Lys	Gly	Thr

Leu Ala His Arg Asp Phe Ser Ala Glu Tyr Glu Glu Asp Gly Lys 3950 3960 5. Phe Glu Gly Leu Gln Glu Trp Glu Gly Lys Ala His Leu Asn Ile 3965 3970 3975 Lys Ser Pro Ala Phe Thr Asp Leu His Leu Arg Tyr Gln Lys Asp 3980 3985 3990 10 Lys Lys Gly Ile Ser Thr Ser Ala Ala Ser Pro Ala Val Gly Thr 3995 4000 4005 15 Val Gly Met Asp Met Asp Glu Asp Asp Phe Ser Lys Trp Asn 4010 4015 20 Phe Tyr Tyr Ser Pro Gln Ser Ser Pro Asp Lys Lys Leu Thr Ile 4025 4030 4035 25 Phe Lys Thr Glu Leu Arg Val Arg Glu Ser Asp Glu Glu Thr Gln 4040 4045 4050 30 Ile Lys Val Asn Trp Glu Glu Glu Ala Ala Ser Gly Leu Leu Thr 4055 4060 4065 Ser Leu Lys Asp Asn Val Pro Lys Ala Thr Gly Val Leu Tyr Asp 35 Tyr Val Asn Lys Tyr His Trp Glu His Thr Gly Leu Thr Leu Arg 4085 4090 4095 40 Glu Val Ser Ser Lys Leu Arg Arg Asn Leu Gln Asn Asn Ala Glu 4100 4105 4110 45 Trp Val Tyr Gln Gly Ala Ile Arg Gln Ile Asp Asp Ile Asp Val 4115 4120 4125 50 Arg Phe Gln Lys Ala Ala Ser Gly Thr Thr Gly Thr Tyr Gln Glu 4130 4135 4140 Trp Lys Asp Lys Ala Gln Asn Leu Tyr Gln Glu Leu Leu Thr Gln 55 Glu Gly Gln Ala Ser Phe Gln Gly Leu Lys Asp Asn Val Phe Asp 4160 4165 4170 60 Gly Leu Val Arg Val Thr Gln Lys Phe His Met Lys Val Lys His 4175 4180 4185 65 Leu Ile Asp Ser Leu Ile Asp Phe Leu Asn Phe Pro Arg Phe Gln 4195 Phe Pro Gly Lys Pro Gly Ile Tyr Thr Arg Glu Glu Leu Cys Thr 4205 70 Met Phe Ile Arg Glu Val Gly Thr Val Leu Ser Gln Val Tyr Ser

Lys Val His Asn Gly Ser Glu Ile Leu Phe Ser Tyr Phe Gln Asp 4235 4240 4245 5 Leu Val Ile Thr Leu Pro Phe Glu Leu Arg Lys His Lys Leu Ile 4250 4255 4260 Asp Val Ile Ser Met Tyr Arg Glu Leu Leu Lys Asp Leu Ser Lys 10 4270 Glu Ala Gln Glu Val Phe Lys Ala Ile Gln Ser Leu Lys Thr Thr 4280 4285 4290 15 Glu Val Leu Arg Asn Leu Gln Asp Leu Leu Gln Phe Ile Phe Gln 4295 4300 4305 20 Leu Ile Glu Asp Asn Ile Lys Gln Leu Lys Glu Met Lys Phe Thr 4310 25 Tyr Leu Ile Asn Tyr Ile Gln Asp Glu Ile Asn Thr Ile Phe Asn 4325 4330 4335 Asp Tyr Ile Pro Tyr Val Phe Lys Leu Leu Lys Glu Asn Leu Cys 30 Leu Asn Leu His Lys Phe Asn Glu Phe Ile Gln Asn Glu Leu Gln 35 4360 Glu Ala Ser Gln Glu. Leu Gln Gln Ile His Gln Tyr Ile Met Ala 4370 4380 40 Leu Arg Glu Glu Tyr Phe Asp Pro Ser Ile Val Gly Trp Thr Val 45 Lys Tyr Tyr Glu Leu Glu Glu Lys Ile Val Ser Leu Ile Lys Asn 4400 4405 4410 Leu Leu Val Ala Leu Lys Asp Phe His Ser Glu Tyr Ile Val Ser 50 4420 Ala Ser Asn Phe Thr Ser Gln Leu Ser Ser Gln Val Glu Gln Phe 55 4430 Leu His Arg Asn Ile Gln Glu Tyr Leu Ser Ile Leu Thr Asp Pro 4445 60 Asp Gly Lys Gly Lys Glu Lys Ile Ala Glu Leu Ser Ala Thr Ala 4460 4465 65 Gln Glu Ile Ile Lys Ser Gln Ala Ile Ala Thr Lys Lys Ile Ile 4480 70 Ser Asp Tyr His Gln Gln Phe Arg Tyr Lys Leu Gln Asp Phe Ser 4490 4495 4500 Asp Gln Leu Ser Asp Tyr Tyr Glu Lys Phe Ile Ala Glu Ser Lys 4505 4510 4515

- Tyr His Thr Phe Leu Ile Arg Leu Ile Asp Leu Ser Ile Gln Asn 4530
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- Ile Asp Ile Tyr Ser Leu Thr Val Asp Ser Arg Val Ser Ser Arg Phe 35
 - Ala His Thr Val Val Thr Ser Arg Val Val Asn Arg Ala Asn Thr Val
- 40 Gin Glu Ala Thr Phe Gin Met Glu Leu Pro Lys Lys Ala Phe Ile Thr
- Asn Phe Ser Met Asn Ile Asp Gly Met Thr Tyr Pro Gly Ile Ile Lys
- Glu Lys Ala Glu Ala Gln Ala Gln Tyr Ser Ala Ala Val Ala Lys Gly
 100 105 50
- Lys Ser Ala Gly Leu Val Lys Ala Thr Gly Arg Asn Met Glu Gln Phe 55
 - Gln Val Ser Val Ser Val Ala Pro Asn Ala Lys Ile Thr Phe Glu Leu
- Val Tyr Glu Glu Leu Leu Lys Arg Arg Leu Gly Val Tyr Glu Leu Leu 145 150 . 155 160
- Leu Lys Val Arg Pro Gln Gln Leu Val Lys His Leu Gln Met Asp Ile 165 170 175
- His Ile Phe Glu Pro Gln Gly Ile Ser Phe Leu Glu Thr Glu Ser Thr 70
- Phe Met Thr Asn Gln Leu Val Asp Ala Leu Thr Thr Trp Gln Asn Lys 200 75
 - Thr Lys Ala His Ile Arg Phe Lys Pro Thr Leu Ser Gln Gln Gln Lys

5 .	~~.	Pr	o Gli	ı Glı	ı Glr	G1u -230	ı Thi	r Val	l Let	ı Ası	235 235	/ Ası	n Le	u Ile	e Ile	e Arg 240
10	Туз	: Ası	o Vai	l Asp	245	Ala	ılle	e Sex	: Gly	7 Gly 250	/ Ser)	: Ile	e Gl	n Ile	Glu 259	ı Asn
15	Gly	Ty	r Phe	≥ Val 260	His	Tyr	Phe	e Ala	265	Glu	ı Gly	Leu	1 Thi	Thr 270		Pro
13	Lys	a Ası	1 Val 275	Val	Phe	Val	Ile	280	Lys	Ser	Gly	' Ser	Met 285	Ser 5	Gly	Arg
20	Lys	11e 290	e Glr	Gln	Thr	Arg	Glu 295	Ala	Leu	Ile	: Lys	Ile 300	Let	Asp	Asp	Leu
25	Ser 305	Pro	Arg	Asp	Gln	Phe 310	Asn	Leu	Ile	Val	Phe 315	Ser	Thr	Glu	Ala	Thr 320
30	Gln	Trp	Arg	Pro	Ser 325	Leu	Val	Pro	Ala	Ser 330	Ala	Glu	Asn	Val	Asn 335	
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45	Arg	Leu 690	Ala	Ile	Leu	Pro	Ala 695	Ser	Ala	Pro	Pro	Ala 700	Thr	Ser	Asn	Pro
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Gly Leu Leu Leu Ser Asp Pro Asp Lys Val Thr Ile Gly Leu Leu Phe Trp Asp Gly Arg Gly Glu Gly Leu Arg Leu Leu Leu Arg Asp Thr 10 Asp Arg Phe Ser Ser His Val Gly Gly Thr Leu Gly Gln Phe Tyr Gln Glu Val Leu Trp Gly Ser Pro Ala Ala Ser Asp Asp Gly Arg Arg Thr 15 Leu Arg Val Gln Gly Asn Asp His Ser Ala Thr Arg Glu Arg Arg Leu 900 905 910 20 Asp Tyr Gln Glu Gly Pro Pro Gly Val Glu Ile Ser Cys Trp Ser Val 915 920 925 25 Glu Leu 930 <210> 43 <211> 1744 <212> PRT <213> Homo sapiens 35 <300> <308> Swiss-Prot/P01028 <309> 1986-07-21 <313> (1) .. (1744) 40 <400> 43 Met Arg Leu Leu Trp Gly Leu Ile Trp Ala Ser Ser Phe Phe Thr Leu 45 Ser Leu Gln Lys Pro Arg Leu Leu Leu Phe Ser Pro Ser Val Val His 20 25 30 Leu Gly Val Pro Leu Ser Val Gly Val Gln Leu Gln Asp Val Pro Arg
35 40 45 Gly Gln Val Val Lys Gly Ser Val Phe Leu Arg Asn Pro Ser Arg Asn 50 55 55 Asn Val Pro Cys Ser Pro Lys Val Asp Phe Thr Leu Ser Ser Glu Arg 60 Asp Phe Ala Leu Leu Ser Leu Gln Val Pro Leu Lys Asp Ala Lys Ser 65 Cys Gly Leu His Gln Leu Leu Arg Gly Pro Glu Val Gln Leu Val Ala 100 105 110

His Ser Pro Trp Leu Lys Asp Ser Leu Ser Arg Thr Thr Asn Ile Gln

Gly Ile Asn Leu Leu Phe Ser Ser Arg Arg Gly His Leu Phe Leu Gln

135

70

	Thr 145	Asp	Gln	Pro	Ile	Ту: 15	r A O	sn i	Pro	Gly	Gln	1	Arg 155	Val	Ar	д Ту	r Ar	g Va:	0
5	Phe	Ala	Leu	Asp	Gln 165	Ly	s M	let .	Arg	Pro	Ser 170	Th	r A	sp '	Thr	Ile	Thr 175	Val	
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	Glu	Pro 210		/ Thr	Tr	o Ly	s	11e 215	Ser	Ala	Ar	g Pl	he s	Ser 220	Asp	Gly	Leu	Glu	
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35	Gly	, Ly	s Pr 27		1 G1	n G	ly	V al	Ala 280	Ty:	r Va	ıl A	rg	Phe	Gly 285	Leu	Lev	Asp	ı
	Glu	1 As 29		у Гу	s Ly	's T	hr	Phe 295	Phe	Ar	g Gl	ly I	eu	Glu 300	Ser	Gln	Thr	Lys	í
40	Le:		al As	n Gl	y G	ln S	er 10	His	Ile	e Se	r Le	eu 9	Ser 315	Lys	Ala	Glu	ı Phe	320	k }
45	As	p A	la Le	eu Gl	.u Ly 32	ys I 25	,eu	Asn	Me	t G1	у I 3	le 3	rhr	Asp	Leu	Gli	33!	y Leu 5	1
50	Ar	g L	eu T	yr Va 34	al A	la <i>I</i>	Ala	Ala	a Il	e II 34	.e G 15	lu :	Ser	Pro	Gl3	(G1)	y Gl	u Met	E
55	G1	u G		la G: 55	lu L	eu !	rhr	Set	7r 36	р Т <u>у</u> О	yr P	he '	Val	Ser	36	r Pr	o Ph	e Se	r
33	Le		sp L 70	eu S	er L	ys '	Thr	Ly:	s Ar 5	g H	is L	eu	Val	Pro 380	G1;	y Al	a Pr	o Ph	е
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65	I	le F	ro V	al L	ys V	/al 105	Ser	: Al	a Th	r V	al 9	Ser 110	Ser	Pro	o Gl	y Se	er Va 41	1 Pr .5	ю.
70	G:	lu (/al (sp 1	le	Glr	ı Gl	n As	n T	hr <i>1</i> 25	qeA	Gly	/ Se	r Gl	y GI 43	in Va	al Se	r
75		le 1		(le 1 135	:le :	Ile	Pro	o G1	n Tl 44	nr I 10	le :	Ser	Glı	ı Le	u Gl 44	.n Le 15	eu Se	er Va	:1

Ser Ala Gly Ser Pro His Pro Ala Ile Ala Arg Leu Thr Val Ala Ala

Pro Pro Ser Gly Gly Pro Gly Phe Leu Ser Ile Glu Arg Pro Asp Ser 465 470 480 Arg Pro Pro Arg Val Gly Asp Thr Leu Asn Leu Asn Leu Arg Ala Val 10 Gly Ser Gly Ala Thr Phe Ser His Tyr Tyr Tyr Met Ile Leu Ser Arg
500 505 510 15 Gly Gln Ile Val Phe Met Asn Arg Glu Pro Lys Arg Thr Leu Thr Ser 515 520 525 Val Ser Val Phe Val Asp His His Leu Ala Pro Ser Phe Tyr Phe Val 530 535 540 20 Ala Phe Tyr Tyr His Gly Asp His Pro Val Ala Asn Ser Leu Arg Val 545 550 555 560 25 Asp Val Gln Ala Gly Ala Cys Glu Gly Lys Leu Glu Leu Ser Val Asp 565 575 30 Gly Ala Lys Gln Tyr Arg Asn Gly Glu Ser Val Lys Leu His Leu Glu
580 35 Thr Asp Ser Leu Ala Leu Val Ala Leu Gly Ala Leu Asp Thr Ala Leu 595 600 605 40 Tyr Ala Ala Gly Ser Lys Ser His Lys Pro Leu Asn Met Gly Lys Val 610 620 Phe Glu Ala Met Asn Ser Tyr Asp Leu Gly Cys Gly Pro Gly Gly 625 635 640 45 Asp Ser Ala Leu Gln Val Phe Gln Ala Ala Gly Leu Ala Phe Ser Asp 645 650 655 50 Gly Asp Gln Trp Thr Leu Ser Arg Lys Arg Leu Ser Cys Pro Lys Glu 660 665 670 55 Lys Thr Thr Arg Lys Lys Arg Asn Val Asn Phe Gln Lys Ala Ile Asn 675 680 685 60 Glu Lys Leu Gly Gln Tyr Ala Ser Pro Thr Ala Lys Arg Cys Cys Gln 690 695 700 Asp Gly Val Thr Arg Leu Pro Met Met Arg Ser Cys Glu Gln Arg Ala 705 710 715 720 65 Ala Arg Val Gln Gln Pro Asp Cys Arg Glu Pro Phe Leu Ser Cys Cys 725 730 735 70 Gln Phe Ala Glu Ser Leu Arg Lys Lys Ser Arg Asp Lys Gly Gln Ala
740 745 750 75 Gly Leu Gln Arg Ala Leu Glu Ile Leu Gln Glu Glu Asp Leu Ile Asp 755 760 765

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55	T	yr V	al A	rg V	al T 80	hr A	la S	er A	sp P	ro L 85	eu As	Tì	nr Le	eu Gl 99	ly S∈ 90	er Glu
60) G	ly A	la L 9	eu \$ 95	er P	ro G	ly G	ly V	al .000	Ala	Ser 1	Leu 1	Leu i	Arg L005	Leu	Pro Arg
6:		ly C	ys 1010	Gly	Glu	Gln	Thr	Met 1015	Ile	Tyr	Leu	Ala	Pro 102	Th: O	r Le	u Ala
U.	,	ala s	Ser 1025	Arg	Tyr	Leu	Asp	Lys 103(Th:	Glu	. Gln	Trp	Ser 103	Th 5	r Le	u Pro
7	0	Pro (Glu 1040	Thr	Lys	Asp.	His	Ala 104	V a:	l Ası) Leu	Ile	Gln 105	Ly O	s Gl	y Tyr
7	5 1		Arg 1055		Gln	Gln	Phe	Arg 106	ь С	s Ala	a Asp	G13	/ Ser 106	т <u>у</u> 55	/r Al	la Ala

Trp Leu Ser Arg Asp Ser Ser Thr Trp Leu Thr Ala Phe Val Leu 1075 Lys Val Leu Ser Leu Ala Gln Glu Gln Val Gly Gly Ser Pro Glu 1085 1090 1095 1085 Lys Leu Gln Glu Thr Ser Asn Trp Leu Leu Ser Gln Gln Gln Ala 1100 1105 1110 10 Asp Gly Ser Phe Gln Asp Pro Cys Pro Val Leu Asp Arg Ser Met 1115 1120 1125 15 Gln Gly Gly Leu Val Gly Asn Asp Glu Thr Val Ala Leu Thr Ala 1130 1140 20 Phe Val Thr Ile Ala Leu His His Gly Leu Ala Val Phe Gln Asp 25 Glu Gly Ala Glu Pro Leu Lys Gln Arg Val Glu Ala Ser Ile Ser 1160 1170 Lys Ala Asn Ser Phe Leu Gly Glu Lys Ala Ser Ala Gly Leu Leu 1175 1180 1185 Gly Ala His Ala Ala Ala Ile Thr Ala Tyr Ala Leu Ser Leu Thr 35 Lys Ala Pro Val Asp Leu Leu Gly Val Ala His Asn Asn Leu Met 1205 1210 1215 40 Ala Met Ala Gln Glu Thr Gly Asp Asn Leu Tyr Trp Gly Ser Val 1220 1230 45 Thr Gly Ser Gln Ser Asn Ala Val Ser Pro Thr Pro Ala Pro Arg 1235 1240 1245 50 Asn Pro Ser Asp Pro Met Pro Gln Ala Pro Ala Leu Trp Ile Glu 1250 1260 Thr Thr Ala Tyr Ala Leu Leu His Leu Leu His Glu Gly Lys 55 Ala Glu Met Ala Asp Gln Ala Ser Ala Trp Leu Thr Arg Gln Gly 1280 1290 60 Ser Phe Gln Gly Gly Phe Arg Ser Thr Gln Asp Thr Val Ile Ala 65 Leu Asp Ala Leu Ser Ala Tyr Trp Ile Ala Ser His Thr Thr Glu Glu Arg Gly Leu Asn Val Thr Leu Ser Ser Thr Gly Arg Asn Gly 1325 1330 1335 Phe Lys Ser His Ala Leu Gln Leu Asn Asn Arg Gln Ile Arg Gly 75 1345

Leu Glu Glu Leu Gln Phe Ser Leu Gly Ser Lys Ile Asn Val Lys Val Gly Gly Asn Ser Lys Gly Thr Leu Lys Val Leu Arg Thr Tyr Asn Val Leu Asp Met Lys Asn Thr Thr Cys Gln Asp Leu Gln 1385 1390 1395 10 Ile Glu Val Thr Val Lys Gly His Val Glu Tyr Thr Met Glu Ala 1400 1410 15 Asn Glu Asp Tyr Glu Asp Tyr Glu Tyr Asp Glu Leu Pro Ala Lys 1415 1420 1425 20 Asp Asp Pro Asp Ala Pro Leu Gln Pro Val Thr Pro Leu Gln Leu 1430 1440 Phe Glu Gly Arg Arg Asn Arg Arg Arg Glu Ala Pro Lys Val 25 Val Glu Glu Glu Ser Arg Val His Tyr Thr Val Cys Ile Trp 1460 1465 1470 30 Arg Asn Gly Lys Val Gly Leu Ser Gly Met Ala Ile Ala Asp Val 1480 35 Thr Leu Leu Ser Gly Phe His Ala Leu Arg Ala Asp Leu Glu Lys 40 Leu Thr Ser Leu Ser Asp Arg Tyr Val Ser His Phe Glu Thr Glu 1505 1510 Gly Pro His Val Leu Leu Tyr Phe Asp Ser Val Pro Thr Ser Arg 45 Glu Cys Val Gly Phe Glu Ala Val Gln Glu Val Pro Val Gly Leu 1535 1540 50 Val Gln Pro Ala Ser Ala Thr Leu Tyr Asp Tyr Tyr Asn Pro Glu 1550 1555 1560 55 Arg Arg Cys Ser Val Phe Tyr Gly Ala Pro Ser Lys Ser Arg Leu 1565 1570 1575 60 Leu Ala Thr Leu Cys Ser Ala Glu Val Cys Gln Cys Ala Glu Gly 1580 1585 1590 Lys Cys Pro Arg Gln Arg Arg Ala Leu Glu Arg Gly Leu Gln Asp 65 Glu Asp Gly Tyr Arg Met Lys Phe Ala Cys Tyr Tyr Pro Arg Val 1610 1615 1620 70 Glu Tyr Gly Phe Gln Val Lys Val Leu Arg Glu Asp Ser Arg Ala 75

Ala Phe Arg Leu Phe Glu Thr Lys Ile Thr Gln Val Leu His Phe

		1640)				164	15				1650	•			
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65	Asn	His	Met	Gly	Asn 85	Val	Thr	Phe	Th	r II 90		ro A	la Ası	n Arg	g Glu 95	ı Phe
70	Lys	Ser	Glu	Lys 100	Gly	Arg	Asn	Lys	Ph 10		al T	hr Va	al Gl	n Ala 110		. Phe

Gly Thr Gln Val Val Glu Lys Val Val Leu Val Ser Leu Gln Ser Gly 115 120 125

Tyr Leu Phe Ile Gln Thr Asp Lys Thr Ile Tyr Thr Pro Gly Ser Thr 130 140

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- Glu Asp Ile Ile Ala Glu Glu Asn Ile Val Ser Arg Ser Glu Phe Pro 755 760 765
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- Gly Ile Ser Thr Lys Leu Met Asn Ile Phe Leu Lys Asp Ser Ile Thr 10 785 790 795 800
- Thr Trp Glu Ile Leu Ala Val Ser Met Ser Asp Lys Lys Gly Ile Cys 805 810 815
- Val Ala Asp Pro Phe Glu Val Thr Val Met Gln Asp Phe Phe Ile Asp 820 825 830
- Leu Arg Leu Pro Tyr Ser Val Val Arg Asn Glu Gln Val Glu Ile Arg 835 840 845
- 25 Ala Val Leu Tyr Asn Tyr Arg Gln Asn Gln Glu Leu Lys Val Arg Val 850 855 860
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- Arg His Gln Gln Thr Val Thr Ile Pro Pro Lys Ser Ser Leu Ser Val 885 890 895
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